

# Product Monograph

**PrFORTAZ**

ceftazidime for injection, USP

**Antibiotic**

GlaxoSmithKline Inc.  
7333 Mississauga Rd  
Mississauga, Ontario  
L5N 6L4

Date of Approval:  
September 14, 2017

Control Number: 206545

*©2017 GSK group of companies or its licensor  
Trademarks are owned by or licensed to the GSK group of companies*

## Product Monograph

### **PrFORTAZ**

ceftazidime for injection, USP

### **Antibiotic**

## **Clinical Pharmacology**

*In vitro* studies indicate that the bactericidal action of ceftazidime, a semisynthetic cephalosporin antibiotic, results from inhibition of bacterial cell wall synthesis.

Ceftazidime has a high affinity for the Penicillin-Binding Protein-3 (PBP-3) and moderate affinity for the PBP-1a of certain Gram negative organisms such as *Escherichia coli* and *Pseudomonas aeruginosa*. The affinity for PBP-1b is much less than that for either PBP-3 or PBP-1a. PBP-3 is involved in the process of cross-wall formation (septation). Binding to this protein results in formation of filaments and eventual death of the bacterium. PBP-1a and PBP-1b are involved in longitudinal wall synthesis (elongation) prior to septation. Binding to these proteins results in spheroplast formation followed by rapid lysis.

Ceftazidime has high affinity for PBP-1 and PBP-2 of *Staphylococcus aureus*. However, the drug's affinity for PBP-3 is very much less in this organism.

## Indications and Clinical Use

FORTAZ (ceftazidime for injection) may be indicated for the treatment of patients with infections caused by susceptible strains of the designated organisms in the following diseases:

### Lower Respiratory Tract Infections

Pneumonia caused by *Pseudomonas aeruginosa*; *Haemophilus influenzae* including ampicillin-resistant strains; *Klebsiella* species; *Enterobacter* species; *Proteus mirabilis*; *Escherichia coli*, *Serratia* species, *Streptococcus pneumoniae*, and *Staphylococcus aureus* (methicillin susceptible) strains.

### Urinary Tract Infections

Caused by *Pseudomonas aeruginosa*; *Enterobacter* species; *Proteus* species (indole positive and negative); *Klebsiella* species, and *Escherichia coli*.

Due to the nature of the underlying conditions which usually predispose patients to *Pseudomonas* infections of the lower respiratory and urinary tracts, a good clinical response accompanied by bacterial eradication may not be achieved despite evidence of *in vitro* sensitivity.

### Skin Structure Infections

Caused by *Pseudomonas aeruginosa*; *Klebsiella* species; *Escherichia coli*; *Proteus mirabilis*; *Enterobacter* species; *Staphylococcus aureus* (methicillin susceptible) strains; and *Streptococcus pyogenes*.

## **Bacteremia/Septicemia**

Caused by *Pseudomonas aeruginosa*; *Klebsiella* species; *Escherichia coli*; *Serratia* species; *Streptococcus pneumoniae*; *Staphylococcus aureus* (methicillin susceptible) strains; and *Staphylococcus epidermidis*.

## **Bone Infections**

Caused by *Pseudomonas aeruginosa*; *Proteus mirabilis*; *Enterobacter* species; and *Staphylococcus aureus* (methicillin susceptible) strains.

## **Peritonitis**

Caused by *Escherichia coli*; *Klebsiella* species; and *Peptostreptococcus* species. Patients infected with *Bacteroides* species have also responded.

## **Meningitis**

Caused by *Haemophilus influenzae* and *Neisseria meningitidis*. FORTAZ has also been used successfully in a limited number of cases of meningitis due to *Pseudomonas aeruginosa*.

## **General**

Specimens for bacteriologic culture should be obtained prior to therapy in order to identify the causative organisms and to determine their susceptibilities to FORTAZ. Therapy may be instituted before results of susceptibility testing are known. However, modification of the treatment may be required once these results become available.

To reduce the development of drug-resistant bacteria and maintain the effectiveness of FORTAZ and other antibacterial drugs, FORTAZ should be used only to treat infections that are proven or strongly suspected to be caused by susceptible bacteria. When

culture and susceptibility information are available, they should be considered in selecting or modifying antibacterial therapy. In the absence of such data, local epidemiology and susceptibility patterns may contribute to the empiric selection of therapy.

## **Contraindications**

FORTAZ (ceftazidime for injection) is contraindicated for patients who have shown hypersensitivity to ceftazidime or the cephalosporin group of antibiotics.

## **Warnings**

Before therapy with FORTAZ (ceftazidime for injection) is instituted, careful enquiry should be made to determine whether the patient has had previous hypersensitivity reactions to ceftazidime, cephalosporins, penicillins, or other drugs. FORTAZ should be administered with caution to any patient who has demonstrated some form of allergy, particularly to drugs. Special care is indicated in patients who have experienced an allergic reaction to penicillins or other beta-lactams. If an allergic reaction to FORTAZ occurs, treatment should be discontinued and standard agents (e.g. epinephrine, antihistamines, corticosteroids) administered as necessary. Elevated levels of ceftazidime in patients with renal insufficiency can lead to convulsions. (see **PRECAUTIONS**).

### ***Clostridium difficile*-Associated Disease**

*Clostridium difficile*-associated disease (CDAD) has been reported with use of many antibacterial agents, including FORTAZ (ceftazidime for injection). CDAD may range in severity from mild diarrhea to fatal colitis. It is important to consider this diagnosis in

patients who present with diarrhea, or symptoms of colitis, pseudomembranous colitis, toxic megacolon, or perforation of colon subsequent to the administration of any antibacterial agent. CDAD has been reported to occur over 2 months after the administration of antibacterial agents.

Treatment with antibacterial agents may alter the normal flora of the colon and may permit overgrowth of *Clostridium difficile*. *C. difficile* produces toxins A and B, which contribute to the development of CDAD. CDAD may cause significant morbidity and mortality. CDAD can be refractory to antimicrobial therapy.

If the diagnosis of CDAD is suspected or confirmed, appropriate therapeutic measures should be initiated. Mild cases of CDAD usually respond to discontinuation of antibacterial agents not directed against *Clostridium difficile*. In moderate to severe cases, consideration should be given to management with fluids and electrolytes, protein supplementation, and treatment with an antibacterial agent clinically effective against *Clostridium difficile*. Surgical evaluation should be instituted as clinically indicated, as surgical intervention may be required in certain severe cases (see ADVERSE REACTIONS).

## **Hemolytic Anemia**

**FORTAZ SHOULD NOT BE USED IN PATIENTS WITH A HISTORY OF CEPHALOSPORIN-ASSOCIATED HEMOLYTIC ANEMIA SINCE THE RECURRENCE OF HEMOLYSIS IS MUCH MORE SEVERE.**

An immune mediated hemolytic anemia has been observed in patients receiving cephalosporin class antibacterials, including FORTAZ. Severe cases of hemolytic anemia, including fatalities, have been reported in both adults and children. If a patient

develops anemia anytime during, or within 2-3 weeks subsequent to the administration of FORTAZ, the diagnosis of a cephalosporin-associated anemia should be considered and the drug discontinued until the etiology is determined.

Patients may benefit from periodic monitoring for signs and symptoms of hemolytic anemia, including measurement of hematological parameters or drug-induced antibody testing, where appropriate (see **ADVERSE REACTIONS**).

## **Susceptibility / Resistance**

### ***Development of Drug-Resistant Bacteria***

Prescribing FORTAZ in the absence of a proven or strongly suspected bacterial infection is unlikely to provide benefit to the patient and risks the development of drug-resistant bacteria.

Development of resistance during the administration of FORTAZ has been observed for *Staphylococcus aureus*, members of the *Enterobacteriaceae* family, *Acinetobacter* species, *Pseudomonas* species, and *Serratia* species.

The prevalence of acquired resistance is geographically and time dependent and for select species may be very high. Local information on resistance and prevalence of extended spectrum beta lactamase (ESBLs) producing organisms is desirable, particularly when treating severe infections.

### ***Potential for Microbial Overgrowth***

Prolonged treatment with FORTAZ may result in the overgrowth of nonsusceptible organisms, including species originally sensitive to the drug. Repeated evaluation of the patient's condition is essential. If superinfection occurs during therapy, appropriate measures should be taken.

## Precautions

### General

FORTAZ (ceftazidime for injection) should be administered with caution to individuals with a history of gastrointestinal disease, particularly colitis.

Patients with impaired renal function (i.e. creatinine clearance of 50 mL/min/1.73 m<sup>2</sup> or less) should be placed on the special dosage schedule for FORTAZ recommended under DOSAGE AND ADMINISTRATION. Normal dosages in these individuals are likely to produce excessive serum concentrations of ceftazidime. Elevated levels of ceftazidime in these patients could lead to convulsions.

Ceftazidime is eliminated via the kidneys, therefore the dosage should be reduced according to the degree of renal impairment. Neurological sequelae have occasionally been reported when the dose has not been reduced appropriately (see Dosage in Impaired Renal Function and see **ADVERSE REACTIONS**).

### Sodium Content

Each 1 g of ceftazidime contains 52 mg of sodium. The sodium content must be taken into account in patients requiring sodium restriction.

### Pregnancy

The safety of FORTAZ in pregnancy has not been established. The use of FORTAZ in pregnant women requires that the likely benefit from the drug be weighed against the possible risk to the mother and fetus.



Reproduction studies have been performed in mice and rats employing ceftazidime doses of up to 25 times those usually administered to humans. These studies have revealed no evidence of impaired fertility or harm of the fetus caused by ceftazidime. Animal reproduction studies, however, are not always predictive of human response.

### **Nursing Mothers**

Ceftazidime is excreted in human milk in low concentrations (3.8 - 5.2 mg/L). The clinical significance of this is unknown, therefore, caution should be exercised when FORTAZ is administered to a nursing mother.

### **Elderly Patients**

Because elderly patients are more likely to have decreased renal function, care should be taken in dose selection, and it may be useful to monitor renal function.

### **Carcinogenesis, Mutagenesis, Impairment of Fertility**

Long-term studies in animals have not been performed to evaluate carcinogenic potential. However, a mouse micronucleus test and an Ames test were both negative for mutagenic effects.

### **Drug Interactions**

The concomitant administration of aminoglycosides and some cephalosporins has caused nephrotoxicity. Although transient elevations of BUN and serum creatinine have been observed in clinical studies, there is no evidence that FORTAZ, when administered alone, is significantly nephrotoxic. However, the effect of administering FORTAZ concomitantly with aminoglycosides is not known. Studies suggest that the concomitant

use of potent diuretics, such as furosemide and ethacrynic acid, may increase the risk of renal toxicity with cephalosporins.

Chloramphenicol is antagonistic *in vitro* with ceftazidime and other cephalosporins. The clinical relevance of this finding is unknown, but if concurrent administration of ceftazidime with chloramphenicol is proposed, the possibility of antagonism should be considered.

In common with other antibiotics, ceftazidime may affect the gut flora, leading to lower estrogen reabsorption and reduced efficacy of combined oral contraceptives.

### **Drug-Laboratory Test Interactions**

Ceftazidime may cause a false positive reaction for glucose in the urine with copper reduction tests (Benedict's or Fehling's solution). As a false negative result may occur in the ferricyanide test, it is recommended that either glucose oxidase or hexokinase method be used to determine blood plasma glucose levels in patients receiving FORTAZ.

Ceftazidime does not interfere in the alkaline picrate assay for creatinine.

A positive Coombs' test has been reported during treatment with cephalosporins. This phenomenon can interfere with cross matching of blood.

## **Adverse Reactions**

The most common adverse effects have been local reactions following intravenous injection, allergic reactions and gastrointestinal reactions. Other adverse effects have been encountered less frequently.

**Local (2.8% of patients)**

Thrombophlebitis or phlebitis and pain with intravenous administration. Pain after intramuscular injection.

**Hypersensitivity (2.7% of patients)**

Pruritus, urticaria, macropapular rash, allergic exanthema, and fever.

**Gastrointestinal (<4% of patients)**

Diarrhea, nausea, vomiting, colitis and abdominal pain. Pseudomembranous colitis has been reported (see **WARNINGS**). Oral thrush has been reported very rarely.

**Central Nervous (<1% of patients)**

Headache, dizziness, hallucinations, and lethargy. There have been reports of neurological sequelae including tremor, myoclonia, convulsions encephalopathy and coma occurring in patients with renal impairment in whom the dose of ceftazidime has not been appropriately reduced.

**Renal (<1% of patients)**

Transient elevations of blood urea, blood urea nitrogen and serum creatinine.

**Hepatic (<4% of patients)**

Transient elevations of serum bilirubin, alkaline phosphatase, LDH, AST (SGOT), ALT (SGPT) and GGT.

**Hematopoietic**

Eosinophilia (3.4%), positive Direct Coombs' Test (5.1%), and with an incidence of <1%: thrombocytosis, transient leukopenia, neutropenia, thrombocytopenia (see **WARNINGS**).

### **Miscellaneous (<1% of patients)**

Blurred vision, flushing, candidiasis, and vaginitis.

## **POST-MARKETING EXPERIENCE WITH FORTAZ**

In addition to adverse events reported during clinical trials, the following events have been identified during clinical practice in patients treated with FORTAZ and were reported spontaneously. Because these reactions were reported voluntarily from a population of uncertain size, it is not possible to reliably estimate their frequency.

### **Blood and lymphatic system disorders**

Lymphocytosis, hemolytic anemia, and agranulocytosis.

### **Immune system disorders**

Anaphylaxis (including bronchospasm and/or hypotension).

### **Nervous system disorders**

Paraesthesia.

### **Gastrointestinal disorders**

Bad taste.

### **Hepatobiliary disorders**

Jaundice.

### **Skin and subcutaneous tissue disorders**

Angioedema, erythema multiforme, Stevens-Johnson syndrome, and toxic epidermal necrolysis.

## Symptoms and Treatment of Overdosage

For management of a suspected drug overdose contact your regional Poison Control Centre.
--

Overdosage of cephalosporins can lead to neurological sequelae including encephalopathy, convulsions and coma. Excessive serum levels of ceftazidime can be reduced by hemodialysis or peritoneal dialysis.

## Dosage and Administration

FORTAZ (ceftazidime for injection) may be administered either intravenously or intramuscularly after reconstitution.

Dosage and route of administration should be determined by severity of infection, susceptibility of the causative organism(s), and condition of the patient. The intravenous route is preferable for patients with septicemia, peritonitis or other severe or life threatening infections, or for patients who may be poor risks because of lowered resistance resulting from such debilitating conditions as malnutrition, trauma, surgery, diabetes, heart failure, or malignancy, particularly if shock is present or pending.

The usual duration of treatment is 7 to 14 days. For *Streptococcal* infections, therapy should be continued for at least 10 days.

### Adults

The recommended daily dosage of FORTAZ is 0.5 to 6 g administered in equally divided doses every 8 to 12 hours (see Table 1).

**TABLE 1**

TYPE OF INFECTION	DAILY DOSE IN GRAMS	FREQUENCY AND ROUTE
uncomplicated pneumonia or skin structure infection	1.5 – 3.0	0.5 – 1.0 g IM or IV q8h
uncomplicated urinary tract infections	0.5	250 mg IM or IV q12h
complicated urinary tract infections	1.0 – 1.5	500 mg IM or IV q8h or q12h
bone infections	4.0	2 g IV q12h
peritonitis or septicemia	6.0	2 g IV q8h
meningitis	6.0	2 g IV q8h

For the treatment of infections caused by *Staphylococcus* species, a dosage of 1 or 2 g administered every 8 hours is recommended. For the treatment of infections (except those confined to the urinary tract) caused by *Enterobacter* species, a dosage of at least 1 g administered every 8 hours is recommended.

## Children

**TABLE 2**

Type of Infection	Age group dosage	Dosage
Infections other than meningitis	1 month – 2 months	25 – 50 mg/kg IV q12h to a maximum of 6 g/day
	2 months – 12 years	30 – 50 mg/kg IV q8h to a maximum of 6 g/day
Meningitis	1 month – 12 years	50 mg/kg IV q8h to a maximum of 6 g/day

The maximum daily dose in children is 6 g.

## Neonates (aged 0-28 days)

In children aged one month or less the recommended dose is 25-50 mg/kg of FORTAZ given twice daily.

Data indicates that half life of ceftazidime in neonates increases with decreasing gestational age and can be 3-4 times that in adults. An adjustment in dosing interval may be necessary with an increasing degree of prematurity. Additionally, clearance may increase rapidly in the first 2-3 weeks of life necessitating a readjustment of dose and/or dosing interval.

### **Use in Elderly**

In acutely ill elderly patients with reduced renal clearance of ceftazidime, the daily dosage should not exceed 3 g.

### **Impaired Hepatic Function**

No adjustment in dosage is required for patients with hepatic dysfunction provided renal function is not impaired (see **PHARMACOLOGY**).

### **Adults with Impaired Renal Function**

Ceftazidime is excreted almost exclusively by glomerular filtration. In patients in whom the glomerular filtration rate (GFR) is less than or equal to 50 mL/min (0.83 mL/s), the dosage of FORTAZ must be reduced to compensate for its slower excretion. After an initial loading dose of 1 g of FORTAZ, a maintenance dosage schedule should be followed (see Table 3).

**TABLE 3 Recommended Maintenance doses of FORTAZ in Renal Insufficiency**

Creatinine Clearance		Recommended Unit dose of FORTAZ		Frequency of Dosing*
mL/min/1.73 m <sup>2</sup>	mL/s/1.73 m <sup>2</sup>	Moderate Infections	Severe Infections	
31- 50	0.51 – 0.83	1 g	1.5 g	q12h
16 – 30	0.26 – 0.50	1 g	1.5 g	q24h
6 -15	0.10 – 0.25	500 mg	750 mg	q24h
<5	<0.09	500 mg	750 mg	q48h

\*If the severity of the infection necessitates an increase in the dosing frequency, serum concentrations of ceftazidime should be used as guidelines.

When only serum creatinine levels are known, the following formulae may be used to estimate creatinine clearance. The serum creatinine must represent a steady state of renal function:

**Males**

$$\text{Creatinine clearance (mL/s)} = \frac{\text{Weight (kg)} \times (140 - \text{age})}{49 \times \text{serum creatinine } (\mu\text{mol/L})}$$

OR

$$\text{Creatinine clearance (mL/min)} = \frac{\text{Weight (kg)} \times (140 - \text{age})}{72 \times \text{serum creatinine (mg/dL)}}$$

**Females**

0.85 X above value.

Mean serum half life of ceftazidime in patients with no kidney function was reduced from a range of 24.0 - 35.4 h between dialysis sessions to a range of 2.8 - 4.6 h during hemodialysis. Therefore a loading dose of 1 g is recommended followed by 0.5 to 1.0 g after each hemodialysis period. Serum concentrations of ceftazidime should be carefully monitored and used as a basis to adjust the dosage.



FORTAZ can also be used in patients undergoing peritoneal dialysis and continuous ambulatory peritoneal dialysis. In such patients, a loading dose of FORTAZ (1 g) is suggested, followed by 500 mg every 24 hours. Serum concentrations of ceftazidime should be carefully monitored and used as a basis to adjust the dosage.

For patients in renal failure receiving low-flux haemofiltration, the dosage as recommended under impaired renal function should be followed. For patients in renal failure receiving continuous arteriovenous haemodialysis or high-flux haemofiltration, 1 g of FORTAZ daily either as a single dose or in divided doses may be administered.

Clinical studies on safety and efficacy of FORTAZ in patients on continuous venovenous hemofiltration (CVVH) and continuous venovenous haemodialysis (CVVHD) have not been conducted. Pharmacokinetic modelling data from a limited number of patients with end stage renal disease suggest that ceftazidime clearance is dependent on the ultrafiltration rate and residual renal function in patients receiving CVVH. However, in patients receiving CVVHD, ceftazidime clearance is dependent on ultrafiltration rate, diluent volume, and residual renal function. Therefore a loading dose of 1 g - 2 g followed by a maintenance dosage of 0.25 - 2.0 g every 12 hours (total daily dose 0.5- 4 g) and 0.5 g -2 g every 12 hours (total daily dose 1- 4 g) may be considered in patients on CVVH or CVVHD, respectively.

A clinical judgement for individual patient dose optimisation should be considered based on severity of the infection, susceptibility of the causative organism and therapeutic monitoring. Dosage should be adjusted to maintain drug levels  $\geq 4$  times the minimum inhibitory concentration (MIC) for gram negative susceptible pathogens.

## **ADMINISTRATION**

### ***Intramuscular***

FORTAZ may be administered by deep intramuscular injection into a large muscle mass such as the upper outer quadrant of the gluteus maximus or vastus lateralis. The maximum dose of FORTAZ should be one (1) gram for a single intramuscular injection.

### ***Intermittent Intravenous Administration***

The reconstituted solution may be slowly injected into the vein over a period of 3 to 5 minutes or given through the tubing of an administration set. During the infusion of the solution containing FORTAZ, the administration of other solutions should be discontinued temporarily.

### ***Continuous Intravenous Infusion***

FORTAZ may also be administered over a longer period of time.

NOTE: If therapy with FORTAZ is carried out in combination with an aminoglycoside antibiotic, each should be administered at different sites because of a physical incompatibility. An aminoglycoside should not be mixed with FORTAZ in the same container.

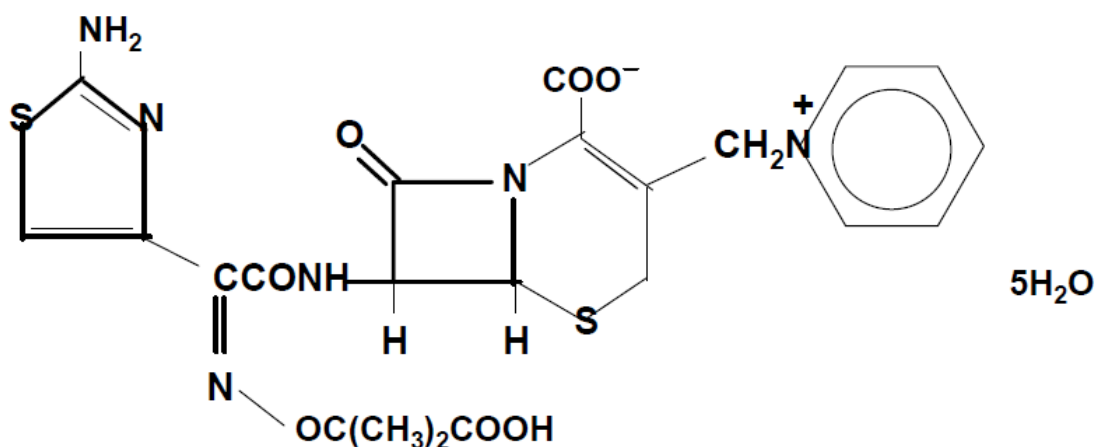
# Pharmaceutical Information

## CHEMISTRY

Proper Name: ceftazidime for injection

Chemical Name: Pyridinium, 1-[[7-[[[(2-amino-4-thiazolyl) [(1-carboxy-1-methylethoxy)imino]acetyl]amino]-2-carboxy-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-en-3-yl]methyl]-, hydroxide, inner salt, pentahydrate, [6R- [6 $\alpha$ , 7 $\beta$ (Z)]]-

Structural Formula:



Molecular Formula: C<sub>22</sub>H<sub>22</sub>N<sub>6</sub>O<sub>7</sub>S<sub>2</sub> · 5H<sub>2</sub>O

Molecular Weight: 636.6 (as pentahydrate)

Description: Ceftazidime pentahydrate is a white to cream-coloured powder. It is soluble in acid, alkali and dimethyl sulfoxide; slightly soluble in water, methanol and dimethylformamide; insoluble in 95% ethanol, ethyl acetate, acetone, 1, -4-dioxan, diethyl ether, toluene, petroleum spirit and chloroform.

## **Composition**

FORTAZ vials contain a mixture of ceftazidime pentahydrate and sodium carbonate.

When constituted, this mixture provides a solution of ceftazidime sodium.

The sodium carbonate at a concentration of 118 mg/g of ceftazidime activity has been admixed to facilitate dissolution. The total sodium content of the mixture is approximately 52 mg (2.3 mEq/g of ceftazidime activity).

Solutions of FORTAZ range in colour from light yellow to amber, depending upon the diluent and volume used. The pH of freshly reconstituted solutions usually ranges from 5.0 to 7.5.

## **RECONSTITUTION**

**CAUTION:** Ensure adequate venting, addition of diluent generates a positive pressure.

### ***For Intramuscular Use***

#### **Solutions for Reconstitution**

Sterile Water for Injection or, if required Bacteriostatic Water for Injection with Benzyl Alcohol or Parabens (not for use in neonates) 0.5 w/v to 1.0% w/v Lidocaine Hydrochloride Injection.

## Reconstitution Table

Vial size	Diluent to be added to Vial	Approximate Available Volume	Approximate Average Concentration
1.0 g	3.0 mL	3.9 mL	280 mg/mL

Shake well until dissolved.

### ***For Intravenous Use***

#### Solution for Reconstitution

Sterile Water for Injection

Reconstitute as follows:

## Reconstitution Table

Vial size	Diluent to be added to Vial	Approximate Available Volume	Approximate Average Concentration
1.0 g	10 mL	10.9 mL	100 mg/mL
2.0 g	10 mL	11.7 mL	175 mg/mL

Shake well until dissolved. The prepared solution may be further diluted to the desired volume with any of the solutions listed under "Solutions for IV Infusion".

### ***For Direct Intravenous Injection***

Reconstitute as directed above.

### ***For Intermittent Intravenous Infusion***

Reconstitute as directed above for 1.0 g and 2.0 g vials of FORTAZ.

### ***For Continuous Intravenous Infusion***

Reconstitute 1.0 g and 2.0 g vials of FORTAZ with 10 mL Sterile Water for Injection.

The appropriate quantity of the reconstituted solution may be added to an intravenous bottle containing any of the solutions listed under "Solutions for IV Infusion".

### **6.0 g Pharmacy Bulk Vial**

THE AVAILABILITY OF THE PHARMACY BULK VIAL IS RESTRICTED TO HOSPITALS WITH A RECOGNIZED INTRAVENOUS ADMIXTURE PROGRAM.

FORTAZ FOR INJECTION does not contain any preservatives. The Pharmacy Bulk Vial is intended for multiple dispensing for intravenous use only, employing a single puncture. Reconstitute with 26 mL Sterile Water for Injection.

### **Reconstitution Table**

<b>Vial size</b>	<b>Diluent to be added to vial</b>	<b>Approximate Available Volume</b>	<b>Approximate Average Concentration</b>
6.0 g	26 mL	30 mL	200 mg/mL

Shake well until dissolved. Following reconstitution with Sterile Water for Injection, the solution should be dispensed and diluted for use within 8 hours at room temperature (not exceeding 25°C). Any unused reconstituted solution should be discarded after 8 hours. The appropriate quantity of the reconstituted solution may be added to an intravenous bottle containing any of the solutions listed below.

### ***Solutions for IV Infusion***

0.9% Sodium Chloride Injection

M/6 Sodium Lactate Injection

Ringers Injection USP

Lactated Ringers Injection USP

5% Dextrose Injection

5% Dextrose and 0.225% Sodium Chloride Injection

5% Dextrose and 0.45% Sodium Chloride Injection

5% Dextrose and 0.9% Sodium Chloride Injection

10% Dextrose Injection

10% Invert Sugar in Water for Injection

Normosol-M in 5% Dextrose Injection

Sterile Water for Injection

### ***Stability of Solutions***

#### ***Storage***

Reconstituted solutions should be administered within 12 hours when stored at room temperature (not exceeding 25°C), and within 48 hours when refrigerated, from the time of reconstitution, both when prepared as bolus injections, IM or IV, and as infusion admixtures with the recommended IV diluents.

#### ***Incompatibility***

FORTAZ should not be added to blood products, protein hydrolysates or amino acids. FORTAZ should not be mixed together with an aminoglycoside. FORTAZ is less stable in Sodium Bicarbonate Injection than in other intravenous fluids, therefore it is not recommended as a diluent. Precipitation has been reported when vancomycin has been added to FORTAZ in solution. Therefore, it would be prudent to flush giving sets and intravenous lines between administration of these two agents.

## Availability of Dosage Forms

### ***FORTAZ for intramuscular or direct intravenous injection***

Vials containing the equivalent of 1 g ceftazidime are available in packs of ten.

### ***FORTAZ for intravenous injection or infusion***

Vials containing the equivalent of 1 g and 2 g ceftazidime are available in packs of ten.

Vials containing the equivalent of 6 g ceftazidime are available in packs of 6.

## STORAGE

FORTAZ in the dry state should be stored below 25°C and protected from light.

## MICROBIOLOGY

### **Mechanism of Action**

Ceftazidime is a bactericidal agent that inhibits bacterial cell wall synthesis following attachment to penicillin binding proteins (PBPs). This results in the interruption of cell wall (peptidoglycan) biosynthesis, which leads to bacterial cell lysis and death.

### **Mechanism of Resistance**

Resistance to ceftazidime is primarily through hydrolysis by beta-lactamase, alteration of penicillin-binding proteins (PBPs), outer membrane impermeability, and presence of bacterial efflux pumps.



## **Spectrum of Activity**

Ceftazidime has been shown to be active against most isolates of the following bacteria, both in vitro and in clinical infections (see **INDICATIONS AND CLINICAL USE**).

### **Gram-positive bacteria**

*Staphylococcus aureus* (methicillin susceptible)

*Streptococcus pneumoniae*<sup>a</sup>

*Streptococcus pyogenes*

### **Gram-negative bacteria**

*Enterobacter* species<sup>b</sup>

*Escherichia coli*<sup>b</sup>

*Haemophilus influenzae* (including ampicillin-resistant strains)

*Klebsiella* species<sup>b</sup>

*Neisseria meningitidis*

*Proteus mirabilis*<sup>b</sup>

*Pseudomonas aeruginosa*<sup>b</sup>

*Serratia* species<sup>b</sup>

<sup>a</sup>Some strains of *S. pneumoniae* are resistant to ceftazidime due to alterations in penicillin-binding proteins.

<sup>b</sup>Most extended spectrum beta-lactamase (ESBL)-producing, AmpC and carbapenemase-producing isolates are resistant to ceftazidime.

The following in vitro data are available, but their clinical significance is unknown. At least 90 percent of the following bacteria exhibit an in vitro minimum inhibitory concentration (MIC) less than or equal to the susceptible breakpoint for ceftazidime.

However, the efficacy of ceftazidime in treating clinical infections due to these bacteria have not been established in adequate and well-controlled clinical trials.

### **Gram-positive bacteria**

Coagulase negative *Staphylococcus* (methicillin susceptible)

*Streptococcus agalactiae*

### **Gram-negative bacteria**

*Burkholderia cepacia*

*Haemophilus parainfluenzae*

*Neisseria gonorrhoeae*

*Pasteurella multocida*

*Providencia* species

*Salmonella* species

*Shigella* species

### **Anaerobic bacteria**

*Peptostreptococcus* species

Ceftazidime is not active in vitro against methicillin-resistant staphylococci, *Enterococcus* species, *Listeria monocytogenes*, *Campylobacter* species, *Clostridium difficile*, *Bacteroides* species, *Chlamydia* species, *Mycoplasma* species and *Legionella* species.

## **Susceptibility Test Methods**

Susceptibility to FORTAZ will vary with geography and time (see **Warnings**). Local susceptibility data should be consulted where available.

### ***Dilution Technique:***

Quantitative methods are used to determine antimicrobial MICs. These MICs provide estimates of the susceptibility of bacteria to antimicrobial compounds. The MICs should be determined using a standardized test method (broth and/or agar). The disc diffusion Interpretive criteria are based on the CLSI M100-S24 interpretive criteria as provided in Table 4.

### ***Diffusion Technique:***

Quantitative methods that require measurement of zone diameters can also provide reproducible estimates of the susceptibility of bacteria to antimicrobial compounds. The zone size provides an estimate of the susceptibility of bacteria to antimicrobial compounds. The zone size should be determined using a standardized test method. These procedures use paper discs impregnated with 30 µg FORTAZ to test the susceptibility of bacteria to ceftazidime. The disc diffusion interpretive criteria are provided in Table 4.

**TABLE 4 Disk and MIC breakpoints for ceftazidime susceptibility testing**

Organism	Zone Diameter Interpretive Criteria* (mm) (30 µg disk)			MIC Interpretive Criteria* (µg/mL)		
	Susceptible	Intermediate	Resistant	Susceptible	Intermediate	Resistant
Enterobacteriaceae	≥21	18-20	≤17	≤4	8	≥16
<i>Hemophilus influenza</i>	≥26	-	-	≤2	-	-
<i>Pseudomonas aeruginosa</i>	≥18	15-17	≤14	≤8	16	≥32
<i>Staphylococcus spp.</i>	Susceptibility may be deduced from testing either cefoxitin or oxacillin.					
<i>Streptococcus pneumonia</i>	Penicillin-susceptible <i>S. pneumoniae</i> can be considered susceptible to ceftazidime					
<i>Streptococcus pyogenes</i>	Penicillin-susceptible <i>S. pyogenes</i> can be considered susceptible to ceftazidime					

\*Interpretive criteria based on CLSI M100-S24 interpretive criteria

A report of “Susceptible” indicates the antimicrobial is likely to inhibit growth of the pathogen if the antimicrobial compound reaches the concentrations at the infection site necessary to inhibit growth of the pathogen. A report of “Intermediate” indicates that the result should be considered equivocal, and, if the bacterium is not fully susceptible to alternative, clinically feasible drugs, the test should be repeated. This category implies possible clinical applicability in body sites where the drug product is physiologically concentrated or in situations where a high dosage of the drug product can be used. This category also provides a buffer zone that prevents small uncontrolled technical factors from causing major discrepancies in interpretation. A report of “Resistant” indicates that the antimicrobial is not likely to inhibit growth of the pathogen if the antimicrobial compound reaches the concentrations usually achievable at the infection site; other therapy should be selected.

### **Quality Control:**

Standard ceftazidime powder should provide the range of MIC values noted in Table 5.

The Quality Control should be performed and evaluated according to the CLSI M100-S24 published QC ranges as provided in Table 5.

**TABLE 5 Disk and MIC QC ranges for ceftazidime susceptibility testing**

<b>QC Strain</b>	<b>Disk Range* (mm) (30 µg disk)</b>	<b>MIC Range* (µg/mL)</b>
<i>Escherichia coli</i> ATCC 25922	25-32	0.06-0.5
<i>Pseudomonas aeruginosa</i> ATCC 27853	22-29	1-4
<i>Staphylococcus aureus</i> ATCC 29213	-	4-16
<i>Staphylococcus aureus</i> ATCC 25923	16-20	-
<i>Haemophilus influenzae</i> ATCC 49247	27-35	0.12-1

\*Disk and MIC QC ranges as published from CLSI M100-S24

## **Pharmacology**

### **Animals**

The secondary pharmacological actions of ceftazidime have been studied in four species: mouse, rat, cat and dog. The possible effects of ceftazidime on the central nervous system, cardiovascular, respiratory and autonomic nervous systems, gastrointestinal tract and in smooth muscle contractions have been determined. The following effects were observed:

**TABLE 6**

<b>Animal</b>	<b>Dose</b>	<b>Effect</b>
cat	1 g/kg, IV	increases or decreases in blood pressure and heart rate; transient increase in respiratory rate and minute volume; slight inhibition of contraction of nictitating membrane
mouse rat	4 g/kg, SC	slight inhibition of gastrointestinal propulsion

## **Human**

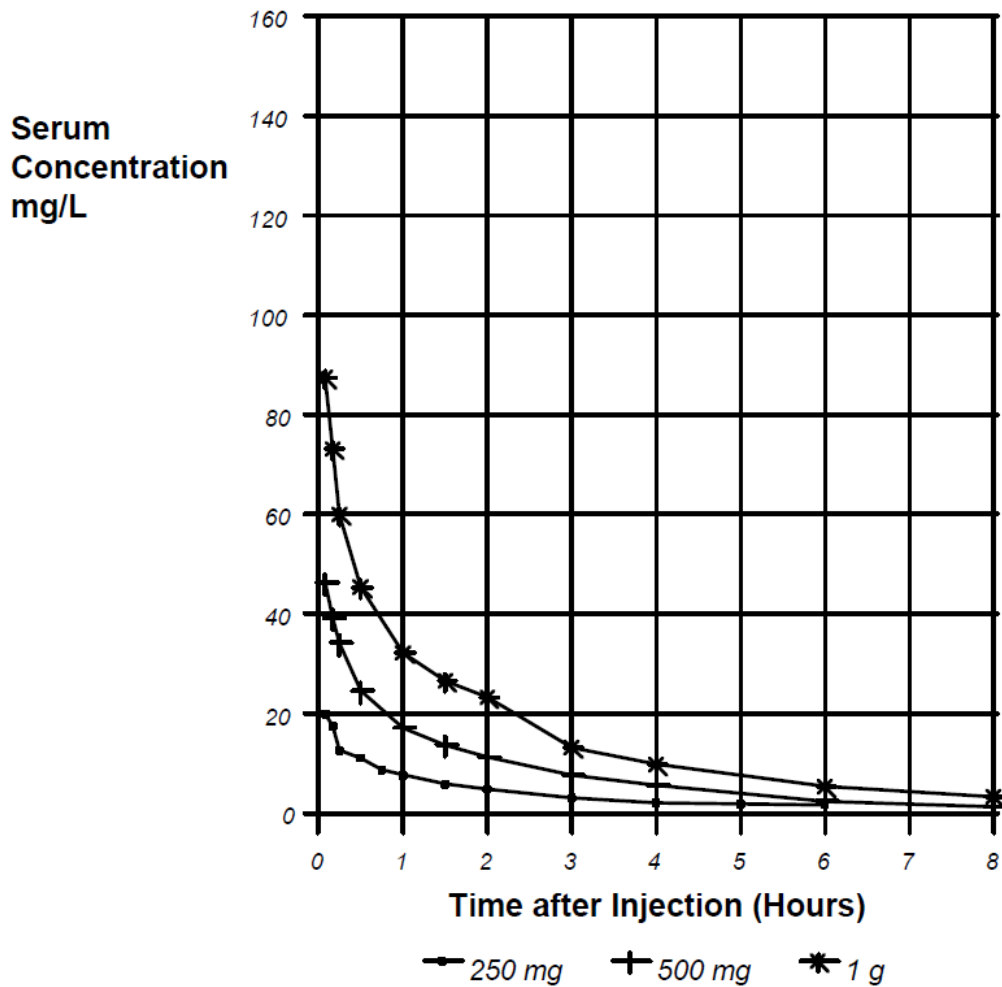
Ceftazidime is poorly absorbed when given orally (e.g. following a 250 mg dose the average urinary recovery was less than 1% of the dose).

### ***Intravenous Administration***

#### **Bolus Injections**

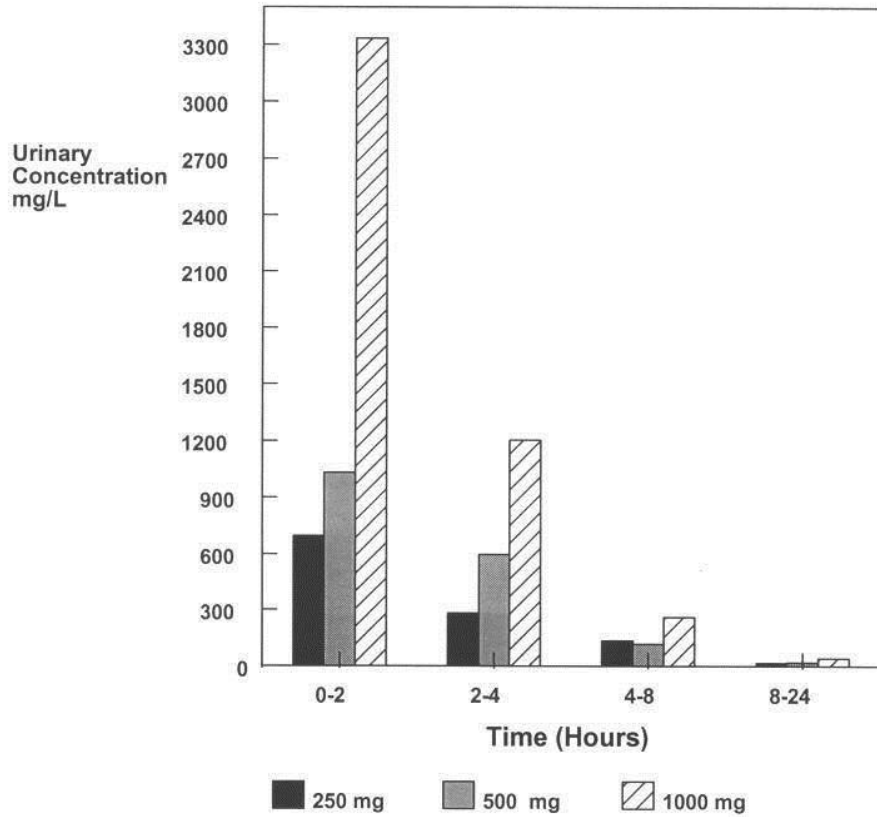
Ceftazidime was administered as single bolus injections (over 1 min) to 22 healthy male volunteers in three doses: 250 mg (6 subjects, mean age 34 years), 500 mg (8 subjects, mean age 33 years) and 1000 mg (8 subjects, mean age 35 years). Serum concentration-time curves follow a biexponential decay (see Figure 1).

**FIGURE 1** Serum concentrations of ceftazidime administered intravenously over 1 minute



Mean urinary recovery of unchanged drug over 24 hours ranged from 77.4 to 85.5% (Table 7) with over 50% being excreted in the first two to four hours. Figure 2 shows urinary concentrations of ceftazidime for various collection intervals following injection. Derived pharmacokinetic indices (based on a two-compartment model) are summarized in Table 7.

**FIGURE 2** Urinary concentrations of ceftazidime after single bolus intravenous injections



No accumulation of drug occurred during repeated administrations of ceftazidime (2 g t.i.d., 10 days). Trough serum level did not increase after dose 2 and urinary recoveries over the first eight hours averaged 81.2% after dose 1 and 76.3% after dose 28.

Pharmacokinetic parameters remained unchanged (see Table 7).



**TABLE 7 Average pharmacokinetic parameters of ceftazidime after IV bolus administration**

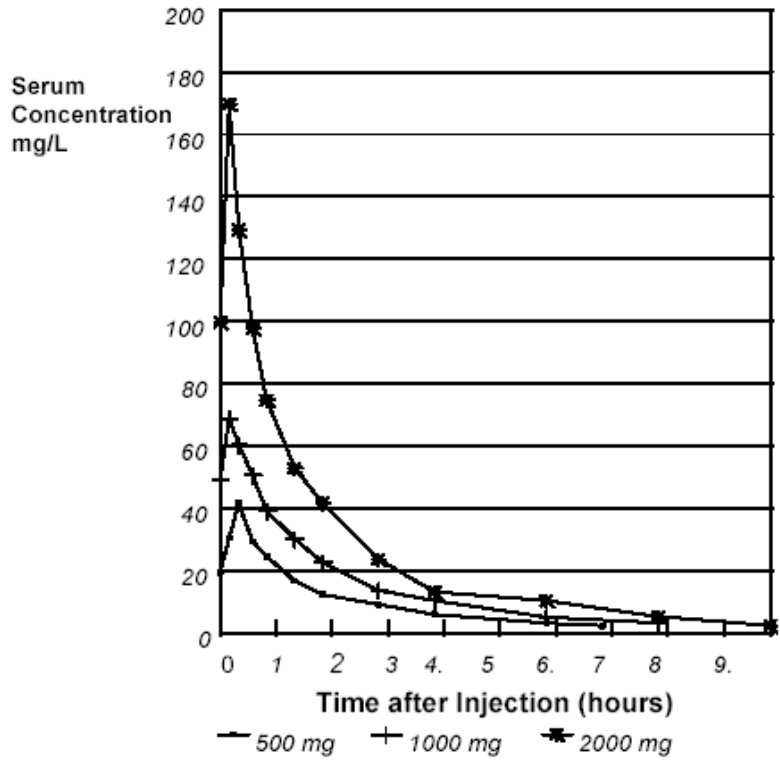
Dose/ Route	Peak Serum Conc (mg/L)	Apparent Volume of distribution (L)	Area under serum level/time curve (mg/L/h)	Serum half-life (h)	Dose recovered in urine to 24 h (%)	Renal Clearance (mL/min)	Plasma Clearance (mL/min)
250 mg IV bolus	28.7	18.9	30.2	1.8	77.4	109	139
500 mg IV bolus	57.6	16.9	71.9	1.9	85.5	100	116
1g IV bolus	119.1	17.1	135.8	1.8	85.1	109	128
2g IV dose 1	182.8	19.7	279.4	1.9	81.2*	102	-
2g IV dose 28	156.7	18.0	274.7	1.7	76.3*	95	-

\* 8h collection only

*Intravenous Infusion*

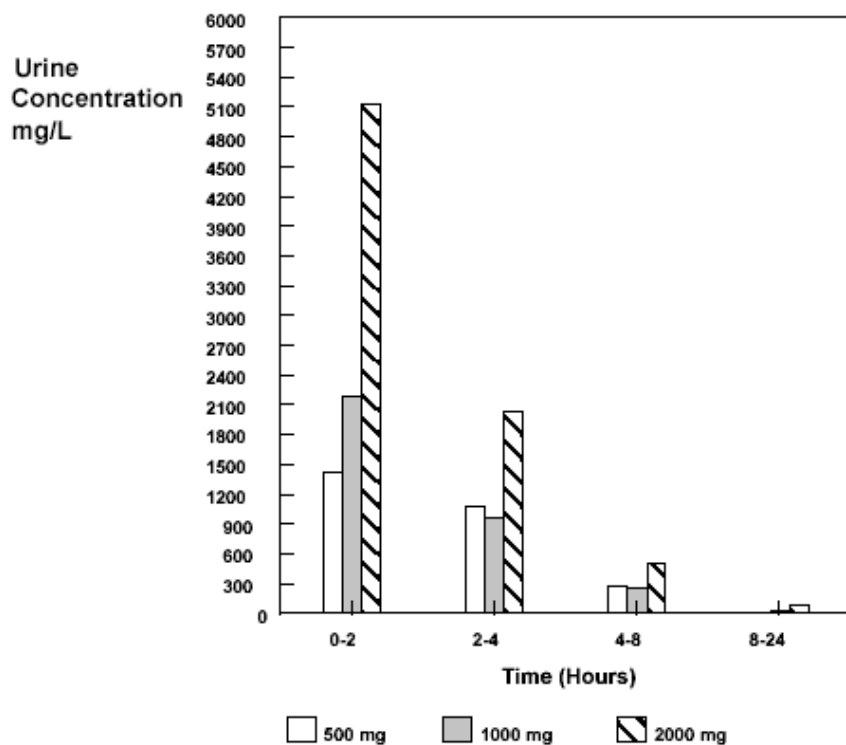
Single intravenous infusions of 500 mg (6 subjects, mean age 35 years), 1000 mg (7 subjects, mean age 33 years) and 2000 mg (7 subjects, mean age 30 years) of ceftazidime were administered over 20 to 30 minutes to normal adult male volunteers. Serum concentration-time curves (Figure 3) follow a biexponential decay.

**FIGURE 3** Serum concentrations of ceftazidime infused intravenously over 20-30 minutes



Mean urinary recovery of unchanged drug over 24 hours ranged from 83.7 to 87.1% (Table 8) with over 50% being excreted in the first two to four hours. Figure 4 shows urinary concentrations of ceftazidime for various collection intervals following infusion. Derived pharmacokinetic indices (based on a two-compartment model) are summarized in Table 8.

**FIGURE 4** Urinary concentrations of ceftazidime after single intravenous infusions over 20-30 minutes



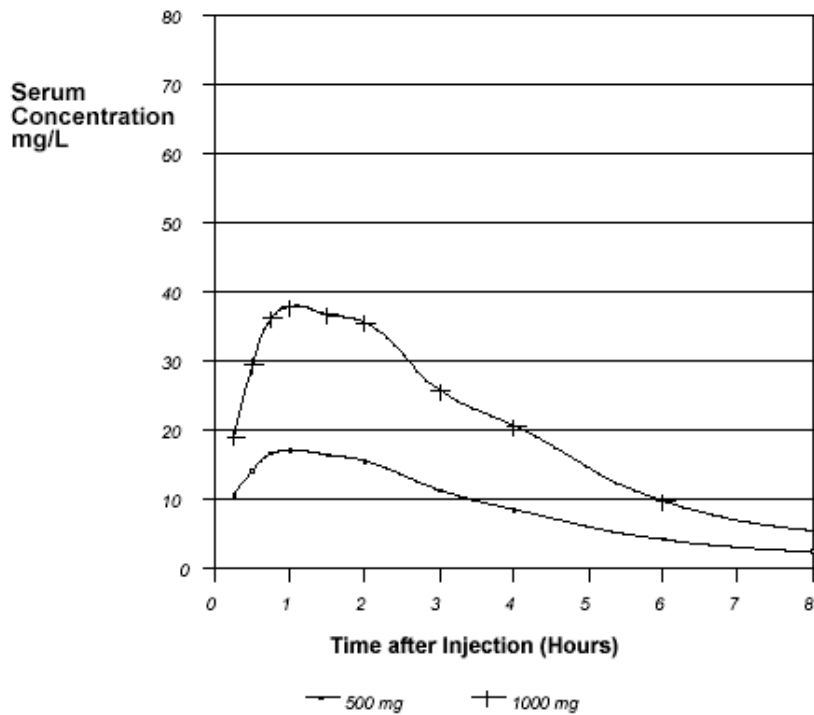
**TABLE 8** Average pharmacokinetic parameters of ceftazidime after IV infusion

Dose/Route	Peak Serum Conc (mg/L)	Apparent Volume of distribution (L)	Area under serum level/time curve (mg/L/h)	Serum half-life (h)	Dose recovered in urine to 24 h (%)	Renal Clearance (mL/min)	Plasma Clearance (mL/min)
500 mg IV infusion	41.5	16.3	82	1.9	86.8	89	102
1g IV infusion	72.1	19.9	143.2	1.9	83.7	98	117
2g IV infusion	170.0	19.9	266.0	1.9	87.1	110	126

Intramuscular Injection:

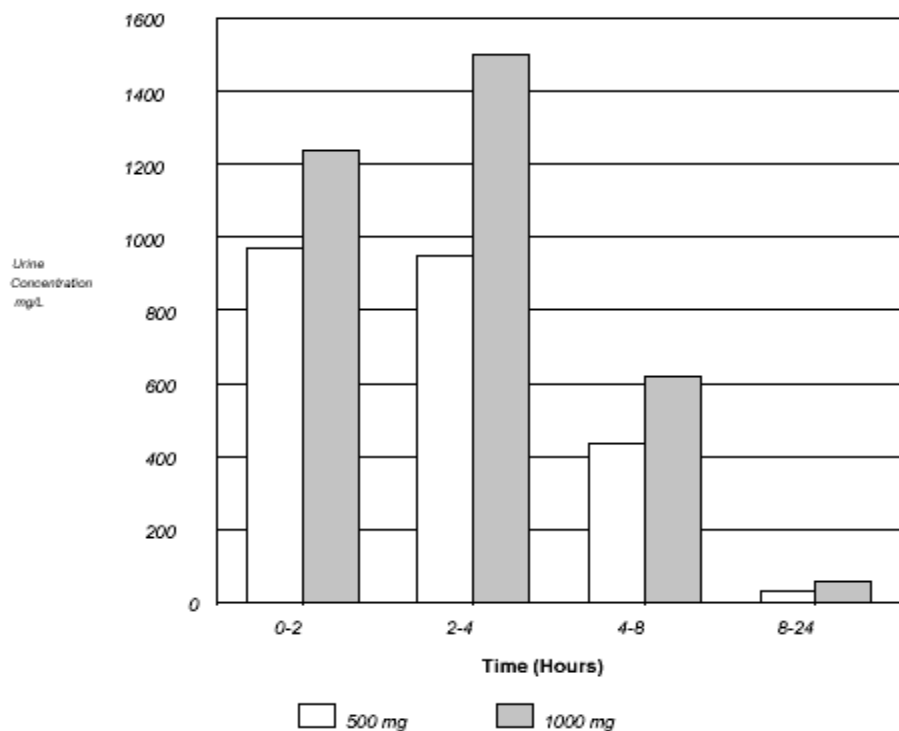
Serum concentration-time curves following intramuscular injection of 500 mg (8 subjects, mean age 32 years) or 1000 mg (8 subjects, mean age 34 years) of ceftazidime is shown in Figure 5.

**FIGURE 5**                    **Serum concentrations of ceftazidime administered intramuscularly**



Mean urinary recovery of ceftazidime over 24 hours ranged from 78.9 to 84.6% (Table 9). Figure 6 shows urinary concentrations of ceftazidime for various collection intervals following injection. Derived pharmacokinetic indices (based on a one-compartment model) are summarized in Table 9.

**FIGURE 6** Urinary concentrations of ceftazidime after single intramuscular injections



**TABLE 9** Average pharmacokinetic parameters of ceftazidime after IM administration

Dose/Route	Peak Serum Conc (mg/L)	Apparent Volume of distribution (L)	Area under serum level/time curve (mg/L/h)	Serum half-life (h)	Dose recovered in urine to 24 h (%)	Renal Clearance (mL/min)	Plasma Clearance (mL/min)
500 mg IM injection	17.4	21.2	79.0	2.2	84.6	90	106
1g IM injection	38.8	16.7	174.7	2.0	78.9	76	97
1g IM dose 1	38.5	16.7	174	2.0	-	-	97
1g IM dose 25	44.0	17.1	186	2.2	-	-	90

No accumulation of drug was noted during repeated intramuscular doses of ceftazidime (1 g, t.i.d., 10 days). Pharmacokinetic parameters remained unchanged (Table 9).

The pharmacokinetic parameters of 1 g of ceftazidime in 1% lidocaine (6 healthy male volunteers, mean age 37 years) did not differ significantly from those obtained without the use of lidocaine.

When ceftazidime was administered to two subjects (750 mg IM) in the recumbent position, average peak serum levels were lower (20.8 mg/L) and serum half-life was longer (2.6 hours) when compared to the two mobile volunteers (36.4 mg/L and 1.8 hours, respectively). The area-under-the-curve was not significantly affected by physical activity.

### ***Excretion and Metabolism***

Ceftazidime is not metabolized. Metabolites were not detected either in the serum by HPLC or in the urine by chromatography or bioautography.

Hepatic clearance (i.e. biliary excretion) accounts for less than 1% of the total clearance of ceftazidime in the presence of normally functioning kidneys.

The mean renal clearance of ceftazidime was 97.6 mL/min (range 76 to 110 mL/min). The calculated plasma clearance of 116.4 mL/min (range 97 - 139 mL/min) indicated nearly complete elimination of ceftazidime by the renal route. Administration of probenecid prior to dosing had no effect on the elimination kinetics of ceftazidime. This

suggested that ceftazidime is eliminated by glomerular filtration and is not actively secreted by renal tubular mechanisms.

### ***Protein Binding***

#### *In vitro*

*In vitro* studies with human serum revealed that 5-23% of ceftazidime is protein bound and is independent of drug concentration.

### ***Tissue and Body Fluid Concentrations***

Therapeutic concentrations of ceftazidime in tissues and body fluids other than serum are presented in Table 10.

**TABLE 10 Ceftazidime Concentration in Body Tissues and Fluids**

TISSUE OR FLUID	DOSE/ROUTE	NO. PATIENTS	TIME OF SAMPLE POST-DOSE	AVERAGE TISSUE OR FLUID LEVEL CONCENTRATION ( $\mu\text{g/mL}$ or $\mu\text{g/g}$ )
Aqueous humour	2 g IV	21	1-3 h	11 $\pm$ 4
Bile	2 g IV	3	90 min	36.4
Blister fluid	1 g IV	7	2-3 h	19.7 $\pm$ 2.3
Bone	2 g IV	5	40 min	31.1 $\pm$ 1.7
Cerebrospinal fluid (inflamed meninges)	2 g q8h IV 2 g q8h IV	5 6	120 min 180 min	9.8 $\pm$ 11.4 9.4 $\pm$ 4.0
Endometrium	2 g IV	6	1-2 h	18.7 $\pm$ 4.7
Fat	2 g IV	39	30-280 min	9.2
Heart Muscle	2 g IV	35	30-280 min	12.7
Lymphatic fluid	1 g IV	7	2-3 h	23.4 $\pm$ 1.2
Myometrium	2 g IV	9	1-2 h	18.9 $\pm$ 4.9
Peritoneal fluid	2 g IV	8	2 h	48.6
Pleural fluid	2 g IV	5	4 h	28 $\pm$ 2
Salpinges	2 g IV	6	1-2 h	18.8 $\pm$ 5.4
Skeletal muscle	2 g IV	35	30-280 min	9.4
Skin	2 g IV	22	30-180 min	6.6
Sputum*	35 mg/kg IV	6	**	2.7
Subcutaneous tissue	2 g IV	2	1-2 h	6.9 $\pm$ 6.3
Synovial fluid	2 g IV	13	2 h	25.6 $\pm$ 1.8

\* Cystic fibrosis patients

\*\* Sputum collected for 8h period

Concentrations of ceftazidime in the breast milk of 11 puerperal women following intravenous administration of 2 g doses every 8 hours for 5 days were determined by bioassay. Mean  $\pm$  S.D. concentrations of ceftazidime averaged 3.8  $\pm$  2.0  $\mu\text{g/mL}$  (before the next dose), 5.2  $\pm$  3.0  $\mu\text{g/mL}$  (1 hour after dosing) and 4.5  $\pm$  1.7  $\mu\text{g/mL}$  (3 hours after



dosing). Excretion of ceftazidime into breast milk remained constant between days 2 and 4 of therapy.

## **Factors Influencing Pharmacokinetics**

### **Sex**

The peripheral comparative volume of distribution was smaller in females (mean  $3.5 \pm 0.5$  L) than in males ( $6.7 \pm 0.6$  L) following intravenous administration (1 g, bolus injection).

Following intramuscular administration (1 g), the time to peak concentration occurred earlier in the men ( $1.0 \pm 0.1$ h - vastus lateralis and  $1.1 \pm 0.1$ h - gluteus maximus) than in women ( $1.3 \pm 0.03$ h and  $1.5 \pm 0.2$ h, respectively). Peak serum concentrations were greater in women ( $37.2 \pm 0.2$  mg/L - vastus lateralis and  $34.0 \pm 2.3$  mg/L - gluteus maximus) than in men ( $29.4 \pm 1.6$  mg/L and  $27.6 \pm 2.3$  mg/L, respectively).

### ***Pregnancy***

Intramuscular injections of at least three doses of ceftazidime (1 g t.i.d.) were administered to 9 pregnant women (mean age 25.6 yr; mean gestational age 20.2 weeks) scheduled for abortion following diagnosis of fetal Cooley's anemia. Amniotic fluid levels of 1.0 - 5.5  $\mu$ g/mL were observed between 2 and 6 hours after dosing. Serum levels of ceftazidime were approximately 50% lower in pregnant than non-pregnant females.

### **Age**

#### **Neonates and Infants**

Two studies were conducted in neonates (aged 0-29 days) which indicated that the serum half-life of ceftazidime in neonates could be 3-4 times that of an adult.

In the first study, 56 neonates (aged less than 29 days) were administered ceftazidime at a dose of 25 mg/kg every 12 hours. The mean serum half-life was 7.57 hours.

In the second study 29 neonates (aged less than 12 days) were dosed with 30-50 mg/kg of ceftazidime every 12 hours had an overall elimination half-life of 4.28 hours. The 30 mg/kg bid dose gave sustained serum levels of ceftazidime throughout the dosing interval and was found to be appropriate for the neonate population.

In another study, conducted in both neonates and infants (1 day to 12 months of age) 53 patients were administered ceftazidime as a single intravenous bolus injection at a mean dose of 31 mg/kg (25.0 - 35.7 mg/kg) in addition to other antimicrobial therapy. Serum levels are presented in Table 11. The mean serum half-life for babies aged 2 months or younger was prolonged ( $4.2 \pm 1.6$ h). Those aged greater than 2 months had a half-life of  $2.0 \pm 0.6$ h.

**TABLE 11**

Age	Serum levels ( $\mu\text{g/mL}$ ) at hrs after dose (mean $\pm$ S.D.)				
	3	5	6	7	9
<2 months (n=30)	54.1 $\pm 28.7$	-	31.2 $\pm 17.9$	-	18.6 $\pm 12.1$
2-12 months (n=23)	26.5 $\pm 10.7$	12.3 $\pm 7.6$	-	6.4 $\pm 6.0$	3.3 $\pm 4.2$

In another study pediatric patients (mean age, 3.5 years) with Gram-negative infections received a single intravenous infusion over 15 minutes of either 15 mg/kg (8 patients) or 50 mg/kg (5 patients) of ceftazidime. Serum levels were measured by bioassay.

Pharmacokinetic data are presented in Table 12.

**TABLE 12 Pharmacokinetic Parameters in Children**

Patient Group	n	Mean Age (Months)	Dose (mg/kg)	Peak Conc. (mg/L)	t <sub>1/2β</sub> (h)	V <sub>d</sub> (L/kg)	C <sub>1</sub> (mL/min/kg)
A	8	22.5	15	37.8	1.65	0.73	5.03
B	5	57.4	50	186.4	1.72	0.52	3.75

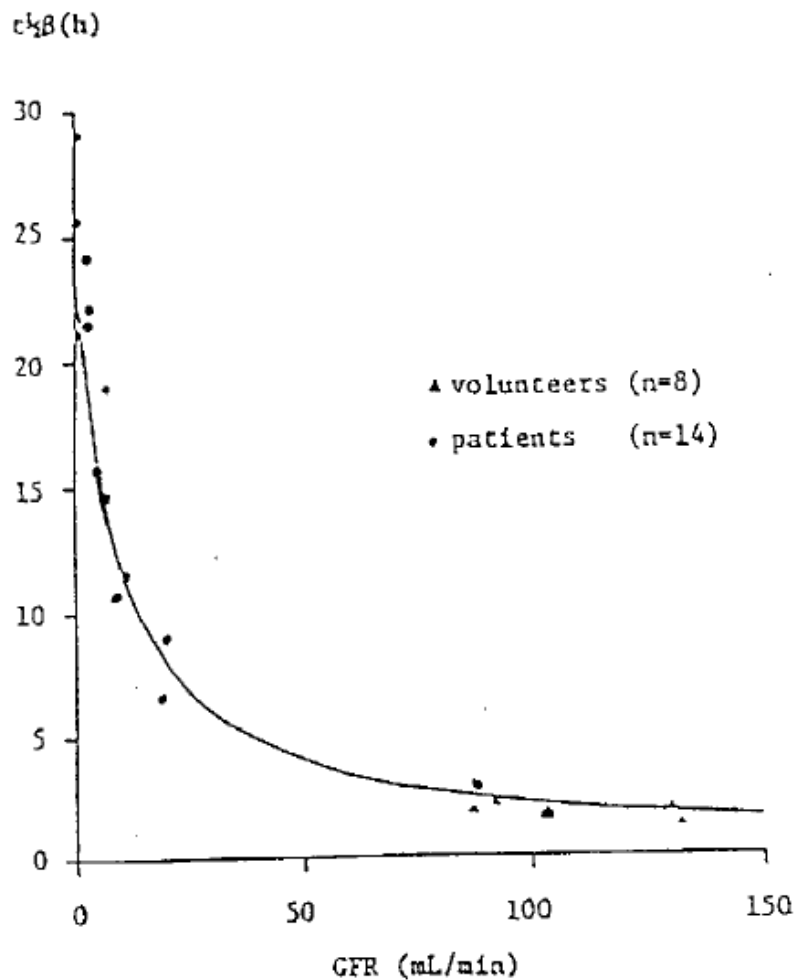
*Elderly*

Ceftazidime, at a dose of 2 g b.i.d., was administered as a bolus intravenous injection to 13 elderly patients with a mean age of 77 years (63 - 83 years) and to 6 younger volunteers (24 - 32 years). A mean serum half-life of 2.9 hours was observed for the elderly patients and 1.75 hours in the young volunteers. The elderly patients were continued on treatment and no accumulation was noted on day 7.

***Impaired Renal Function***

The relationship between serum elimination half-life and glomerular filtration rate (GFR) is curvilinear. The half-life increases steeply at GFR's less-than 50 mL/min/1.73 m<sup>2</sup> (see Figure 7).

**FIGURE 7** The relationship between serum elimination half-life and glomerular filtration rate



The pharmacokinetic parameters obtained following intravenous administration of a 1 g bolus dose of ceftazidime to 14 patients (mean age 49 years) with severely impaired renal function and those from 8 healthy volunteers (mean age 35 years) are given in Table 13.

**TABLE 13 Mean Pharmacokinetic Parameter after 1 g Ceftazidime IV**

Group	C <sub>0</sub> (mg/L)	AUC <sub>T</sub> (mg/h/L)	β (h <sup>-1</sup> )	t <sub>1/2β</sub> (h)	V <sub>dβ</sub> (L)	U <sub>R</sub> (%)	GFR (mL/min)
Volunteers (8)	108	152	0.378	1.9	17.8	88	115
Patients (8)	70	1360	0.061	16.7	19.2	24	12
Patients on Dialysis (6)	82	292	0.176	4.6	22.2	0	--

C<sub>0</sub> = Fictive serum concentration at time zero

AUC<sub>T</sub> = Area under the serum concentration/time curve to infinity

β = Serum elimination rate constant

t<sub>1/2β</sub> = Serum half-life

V<sub>dβ</sub> = Volume of distribution during the post-distributive phase

U<sub>R</sub> = Urinary recovery over 24 h

Mean maximum urine levels ranged between 0.2 g/L in patients with a GFR of <5 mL/min to 0.8 g/L with a GFR of 88 mL/min.

In another study, six normal volunteers and four end-stage renal disease (ESRD) patients on hemodialysis were administered a single 1 g IV dose of ceftazidime. The apparent volumes of distribution were similar in both groups. The terminal half-life in the normal subjects ranged from 1.3 to 1.7 hours, while in the ESRD patients it ranged from 25.5 to 35.4 hours. Dialysis clearance ranged from 27 to 50 mL/min, while the total body clearance in the normals ranged from 98 to 184 mL/min. In another study single bolus doses of ceftazidime (15 mg/kg IV) were administered to 5 normal volunteers and 19 uremic patients (See Table 14). Four of the latter patients received an additional dose during hemodialysis.

**TABLE 14 Pharmacokinetic parameters of ceftazidime in healthy volunteers and in patients with impaired renal function**

Group (n)	Cl <sub>cr</sub> (mL/min)	t <sub>1/2β</sub> (h) (mean ± S.D.)	V <sub>D</sub> (L)
I - Healthy Volunteers (5)	>80	1.5 ± 0.2	17.8 ± 1.2
II - Uremic Patients (5)	30-80	3.6 ± 0.7	17.5 ± 3.1
III - Uremic Patients (6)	13-29	9.0 ± 1.0	16.1 ± 3.4
IV - Uremic Patients (4)	2-12	16.1 ± 4.0	19.1 ± 8.4
V - Hemodialysis Patients (4) - during dialysis - between dialysis sessions	---	2.8 ± 0.2 25.5 ± 4.6	---

Cl<sub>cr</sub> = creatinine clearance    V<sub>D</sub> = volume of distribution

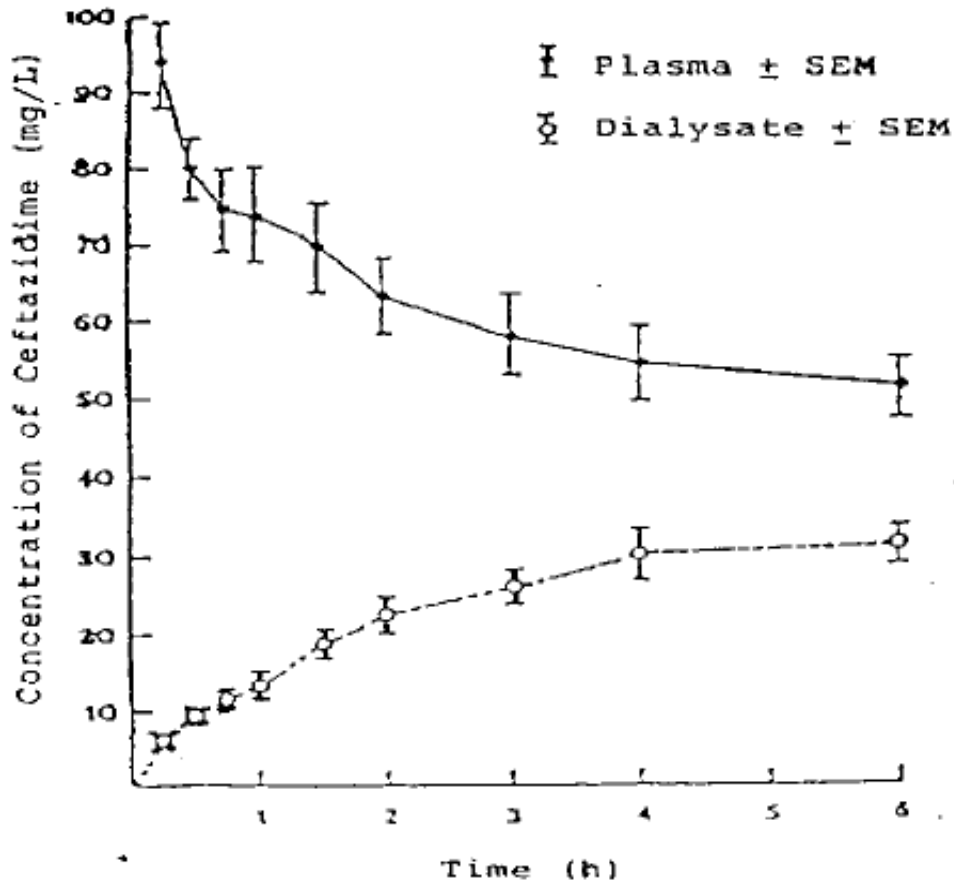
The pharmacokinetics of ceftazidime were studied in 12 patients with end-stage chronic renal failure during peritoneal dialysis. Mean serum levels (mg/L) following the IV administration of ceftazidime (1 g) to 5 patients at 0.25, 2 and 12 hours after starting peritoneal dialysis were 50.6 ± 11.2, 35.6 ± 3.7 and 22.7 ± 7.9 respectively. The mean serum half-life during and after peritoneal dialysis was 8.7 ± 3.1 hours and 26.9 ± 11 hours respectively.

Four patients were administered ceftazidime (1 g) via an intraperitoneal catheter. The mean serum levels (mg/L) at 0.25, 2 and 8 hours were 14.2 ± 3.1, 40 ± 3.1 and 32.5 ± 6.4 respectively.

Five male and one female patient undergoing continuous ambulatory peritoneal dialysis (CAPD) were administered 1 g ceftazidime. Two litres of dialysis fluid were used every six hours. The mean concentrations of ceftazidime in plasma and dialysate are shown in Figure 8. Using a dwell time of 4 to 6 hours, approximately 10% of a dose of ceftazidime

is removed. The data indicate that the half-life of ceftazidime is reduced to approximately 12 hours.

**FIGURE 8** Mean ceftazidime levels in plasma and dialysate of CAPD patients



### **Cystic Fibrosis**

The pharmacokinetics of an intravenous infusion (20 min) of 50 mg/kg ceftazidime were studied in 10 patients ( $20.8 \pm 4.8$  yr, 4 female, 6 males) with cystic fibrosis and 10 normal volunteers ( $21.6 \pm 1.9$  yr, 3 females, 7 males). Serum elimination half-lives were  $1.76 \pm 0.21$  h in controls and  $1.50 \pm 0.19$  h in cystic fibrotics. Total body clearance was 41.9% greater in the cystic fibrosis group ( $142.4 \pm 16.9$  mL/min/1.73 m<sup>2</sup>) compared to

controls ( $100.5 \pm 10.3$  mL/min/ $1.73$  m<sup>2</sup>). Although the fraction of the dose recovered in urine was the same in each group, renal clearance was 40.9% greater in patients with cystic fibrosis ( $130.1 \pm 11.4$  and  $92.7 \pm 11.6$  mL/min/ $1.73$  m<sup>2</sup> respectively).

The mechanisms responsible for the altered renal clearance of ceftazidime in cystic fibrotic patients is not known.

## Toxicology

### Acute Toxicity

TABLE 15

ANIMAL	AGE	SEX	ROUTE	DOSES (g/kg)	ANIMALS /DOSE	LENGTH OF OBSERVATION	LD <sub>50</sub> (g/kg)
mouse	3 days	M	IP	2.1,3.0,4.2,5.9,8.2	8	18 days	4.6 ± 0.6
		F	IP	2.1,3.0,4.2,5.9,8.2,11.5	8	18 days	6.1 ± 0.9
	14 days	M	IP	3.6,4.3,5.2,6.2,7.4,8.9	8	14 days	4.9 ± 0.3
		F	IP	3.6,4.3,5.2,6.2,7.4,8.9	8	14 days	4.8 ± 0.2
	21 days	M	IP	4.7,5.7,6.8,8.2,9.8,11.8	8	14 days	9.0 ± 0.8
		F	IP	4.7,5.7,6.8,8.2,9.8,11.8	8	14 days	8.4 ± 0.6
	adult	M	IV	5.0,6.25,7.5,8.8,10.0	5	14 days	7.0 ± 1.1
		F	IV	5.0,5.6,6.25,6.9,7.5,8.8	5	14 days	6.3 ± 0.6
rat	3 days	M	IP	3.9,4.6,5.6,6.7,8.0,9.6	8	14 days	5.7 ± 0.4
			SC	4.8,5.8,6.9	8	14 days	6
		F	IP	3.9,4.6,5.6,6.7	8	14 days	5.7 ± 0.4
	14 days	M	SC	4.8,5.8,6.9,8.3	8	14 days	6
			IP	4.2,5.0,6.0,7.3,8.7,10.5	8	14 days	5.9 ± 0.6
		F	SC	4.7,5.6,6.7,8.1,9.7,11.6	8	14 days	6.6 ± 0.4
	21 days	M	IP	3.5,4.2,5.0,6.0,7.3,8.7	8	14 days	5.8 ± 0.4
			SC	4.7,5.6,6.7,8.1,9.7,11.6	8	14 days	7.2 ± 0.4
			IP	5.6,6.7,8.1,9.7	8	14 days	7.5 ± 0.4
		F	SC	8.1,9.7,11.7,14.0,16.8	8	14 days	11.9 ± 0.8
			IP	5.6,6.7,8.1,9.7	8	14 days	7.0
			SC	8.1,9.7,11.7,14.0,13.8	8	14 days	12.2 ± 0.7

All deaths in mice occurred within 24 hours after an intravenous dose or within 6 hours following an intraperitoneal dose. Toxic signs consisted of purplish colouration of the skin, immobility and bradypnea alternating with jumping and convulsions. Survivors exhibited no abnormal signs or symptoms at 24 hours following the test dose. Post-mortem examinations revealed meningorrhagia, especially in the cerebellum, and pulmonary congestion.



Death in rats occurred within 24 hours following subcutaneous injection and 6 hours following intraperitoneal dosing. Purplish colouration and reduction in elasticity of the skin, bradypnea, corneal opacity, piloerection, and immobility followed by jumping and convulsions were observed. All survivors appeared normal by 48 hours post-dose. Post-mortem examinations revealed meningorrhagia, pulmonary congestion, splenic cysts and spots, and, in rats who received IP dosing, dilation of the caecum with large amount of contents.

Groups of 6 rats of each sex were given a single intravenous or subcutaneous dose of ceftazidime (5 g/kg). One female died immediately after IV injection while the remaining animals survived in good condition. All animals were sacrificed 7 days post-dose. Post-mortem examinations revealed mild renal damage in the form of paling and slight dilation of some renal tubules containing cellular debris.

A single IV dose of ceftazidime (5 g/kg) was administered to 6-month-old Beagle dogs (2/sex), which survived in good condition except for intermittent emesis and transient tachycardia. The animals were sacrificed 7 days post-dose for extensive histological examinations, but no pathological findings were noted.

## Subchronic and Chronic Toxicity

TABLE 16

ANIMAL	AGES*	ROUTE	DOSES (g/kg/day)	ANIMALS /DOSE**	DURATION OF	
					TREATMENT	RECOVERY
rat	8 - 13 wk	IV	0.0,0.1,0.3,0.9,2.7,8.1	12	30 days	14 days
rat	8 - 13 wk	SC	0.0,0.1,0.3,0.9,2.7,8.1	12	30 days	14 days
rat	unavailable	IM	0.0,0.1,0.3,0.9	20	12 weeks	--
rat	6 - 7 wk	SC	0.0,0.1,0.3,0.9,2.7	20	28 weeks	59 days
rat	5 - 6 wk	SC	0.0,0.1,0.5,2.5	20	27 weeks	21 days
dog	5 - 18 mo	IM	0.00,0.06,0.18,0.54	2 or 4	30-32 days	22-23 days
dog	5 - 18 mo	IV	0.00,0.06,0.18,0.54	4 or 8	30-32 days	22-23 days
dog	5 - 18 mo	SC	0.00,0.06,0.18,0.54	2 or 4	30-32 days	22-23 days
dog	3 wk	IV	0.0,0.1,0.3,1.0	14	35 days	35 days
dog	8 - 10 mo	IM	0.000,0.125,0.250,0.500	6	13 weeks	--
dog	16-27 wk	IV	0.000,0.085,0.255,0.595, 0.85	8	28 weeks	3 weeks

\* Ages at commencement of treatment.

\*\* Each dosage group was composed of equal numbers of males and females.

Rat: 30 day study, intravenous and subcutaneous

All rats given 8.1 g/kg IV and 2 of 12 given 2.7 g/kg died within 10 minutes in convulsive shock and were found to have dilation of proximal and distal convoluted renal tubules.

The 8.1 g/kg dose given sc was tolerated by females for the entire 30 day treatment period; males, however, had to be sacrificed after their third dose, and were found to have coagulative necrosis of 50 to 85% of the proximal convoluted tubules, but no other organ pathology.

Toxicity in survivors was similar by either route of administration. The following biochemical and morphological changes were observed in survivors during or immediately following the treatment period: decreases in serum transaminases, protein (8.1 g/kg/day dose), alkaline phosphatase, calcium, and triglycerides; increases in serum sodium, potassium, inorganic phosphorus, protein (<8.1 g/kg/day doses), and

cholesterol; increased weights of liver, kidney, spleen, ovaries, and adrenals; thymus involution; neutrophilia, lymphocytosis, and normocytic normochromic anemia; and increased urinary volume and output of epithelial cells, protein, and electrolytes. All of these abnormalities regressed during the recovery period.

Post-mortem examination of both treatment mortalities and sacrificed survivors of the 8.1 g/kg/day regimen revealed pulmonary edema, subpleural hemorrhages, fatty change in liver cells, and dilation and fluid content of renal tubules.

Rat: 12 week study, intramuscular

All animals survived treatment and no abnormal physical or behavioural symptoms were observed. The injections were well tolerated at the IM sites.

The following statistically significant changes in laboratory parameters which, nevertheless, still fell within the normal range occurred in the 0.9 g/kg/day group: erythrocyte counts increased in females and decreased in males; decreases in serum alkaline phosphatase, SGPT, hematocrit, and hemoglobin; increases in serum creatinine, bilirubin, potassium, BUN, and SGOT; and inconsistent changes in lymphocyte and neutrophil counts.

The following laboratory abnormalities were observed in the 0.3 and 0.9 g/kg/day groups: increases in serum cholesterol; inconsistent changes in serum proteins; and increases in urinary volume and pH and decreases in specific gravity.

Rat: 28 week study, subcutaneous

One male from the 2.7 g/kg/day group was killed on Day 95 for investigation of suspected hepatotoxicity. Post-mortem examination revealed splenomegaly and hepatic

fibroplasia. A female from the 0.9 g/kg/day group died on Day 183 and was found to have congestion of the lung, thymus, liver and kidney. All other rats survived the entire treatment period. Adverse effects noted in the 2.7 g/kg/day group were local irritation, loose feces, lethargy, decreased weight gain, and a general loss of condition characterized by a rough sticky coat, dirty tail, irregular thickening of the skin, and increased aggressiveness.

Observed laboratory abnormalities in the 0.9 and 2.7 g/kg/day groups were decreases in serum hemoglobin, packed cell volume, and erythrocyte counts; increases in neutrophil, lymphocyte, and platelet counts; increases in prothrombin time; decreases in serum albumin, triglycerides, SGOT, SGPT, and alkaline phosphatase; increases in serum cholesterol and bilirubin; hematuria, bacteriuria, and increases in urinary volume and protein output. Post-mortem examinations revealed increased weights of liver, kidneys, spleen, and adrenals (in females), fibrosis around the central veins of the liver, hemorrhage and fibroplasia at the injection site, and salivary gland edema. Abnormalities which did not regress during the recovery period were increased weights of liver, spleen, adrenals (in females), and kidneys (in males).

Rat: 27 week study, subcutaneous

Environmental control failure resulted in the normal temperatures of 18 – 22°C being exceeded, and rats being exposed to temperatures as high as 29°C.

The toxicity of ceftazidime in heat stressed animals was much higher in females than in males. All females in the 2.5 g/kg/day group died or had to be sacrificed after 8 to 12 weeks of treatment and were found to have extensive centrilobular liver necrosis, and in some, subendocardial fibrosis of the left ventricle. Although males survived the

2.5 g/kg/day dosage regimen, they also showed hepatic changes including fibrosis.

Animals of both sexes in the high dose group were found to have dilation of renal tubules with casts and debris.

Changes in laboratory parameters in both sexes were decreased plasma enzyme activities and hypercholesterolemia at 0.1 g/kg/day or more, increased urinary protein at 0.5 g/kg/day or more, and increased BUN, hyperkalemia, hypoglycemia, hypochromic macrocytic anemia, leukocytosis, thrombocytosis, and increased diuresis at 2.5 g/kg/day.

Dog: 30 to 32 days study, intravenous and subcutaneous/intramuscular

All beagles survived the treatment in good general condition apart from an erythematous skin condition which developed in six dogs, including one control. The IM and SC injections produced dose-related transient pain and irritation which varied from mild to severe. The IV injections caused no apparent local effects but were associated with a dose-related incidence of emesis.

Observed abnormalities consisted of increased total iron binding capacity in males at 0.18 g/kg/day, and increases in relative liver weight, hypoglycemia (in females), and hypertriglyceridemia at 0.54 g/kg/day. Post-mortem examinations revealed no abnormal pathology.

Infant Dog: 35 day study, intravenous

All beagles survived the treatment in good condition. Observed abnormalities consisted of salivation, emesis, and loose feces in the 0.3 and 1.0 g/kg/day groups. These symptoms regressed during the recovery period.

A tendency toward decreased SGOT levels was noted in the 1.0 kg/day group, but the mean change was not statistically significant. No other laboratory abnormalities were observed. Post-mortem findings were also negative.

Dog: 13 week study, intramuscular

All beagles survived the treatment in good condition. The injections were well tolerated at the IM site. No physical or behavioral abnormalities were observed.

The following hematological changes were noted in the 0.25 and 0.50 g/kg/day groups: decreased serum hemoglobin, hematocrit, lymphocytes, and platelets, and prolonged prothrombin time. These changes were statistically significant but the values remained within the normal range.

Other laboratory findings were increases in total serum cholesterol and BUN. Post-mortem examinations revealed protein casts in the lumen of renal tubules in 6 males, including 2 out of the 3 male controls.

Dog: 28 week study, intravenous

Two beagles were sacrificed during the study and found to have, respectively, a cerebellar lesion and polyarteritis. All other dogs survived the study in satisfactory condition. Adverse effects of treatment were discomfort during injection, and a dose-related incidence of salivation and vomiting. Laboratory abnormalities were generally confined to the 0.595 and 0.850 g/kg/day groups and consisted of decreases in serum gamma-globulin and SGPT, and increases in cholesterol, albumin, and total protein. Post-mortem examinations revealed hepatomegaly, injection phlebitis, proteinaceous droplets in proximal convoluted tubular cells, and infiltration of the prostate.

## Nephrotoxicity Studies

TABLE 17

ANIMAL	SEX	ROUTE	CEFTAZIDIM E DOSES (g/kg/day)	ANIMA LS /DOSE	CONCURRENT DRUG AND DOSE	COMPARATIVE DRUG AND DOSE	DURATION OF	
							TREATMENT	OBSERVATION
mouse	F	SC	0,4,6,8,10	10	--	cephaloridine 1.1 g/kg	1 dose	48 hr
	F	SC	0,10	10	furosemide 50 mg/kg	cephaloridine 1.1 g/kg	1 dose	48 hr
	F	SC	0,10	10	probenecid 100 g/kg	cephaloridine 1.1 g/kg	1 dose	48 hr
	F	SC	0,10	10	--	cephaloridine 1.1 g/kg	1 dose	1-7 days
rat	M	SC	0,4	5	--	cephaloridine 2 g/kg	1 dose	1-7 days
	M	SC	0,2,4,6,8,10	6	--	cephaloridine 2 g/kg	1 dose	48 hr
	M	SC	0,4	6	--	cefuroxime 4 g/kg	1 dose	48 hr
	M	SC	0,4	6	gentamicin 35 mg/kg	--	1 dose	48 hr
	M	SC	0,4	6	furosemide 100 mg/kg	cephaloridine 2 g/kg	1 dose	48 hr
	M	SC	0,4	6	probenecid 100 mg/kg	cephaloridine 2 g/kg	1 dose	48 hr
	M	SC	0,4	10	--	--	10 days	10 days
	M	SC	0,4	6	gentamicin 35 mg/kg	--	10 days	24 hr
	M	SC	0,4	6	amikacin 250 mg/kg/day	--	10 days	24 hr
	M	SC	0,4	6	tobramycin 60 mg/kg/day	--	10 days	24 hr
rabbit	M	IM	0,0,0,5	6	--	cephaloridine 0.14 g/kg	1 dose	48 hr
	F	SC	0,0,0,4,0,8	4	--	cephaloridine 0.2 g/kg	1 dose	48 hr
	F	SC	0,0,0,4,0,8	4	--	cefazolin 0.4 g/kg	1 dose	48 hr
	F	SC	0,0,0,4,0,8	4	--	cefazolin 0.8 g/kg	1 dose	48 hr

In female mice, a single sc dose of ceftazidime 6 g/kg resulted in no evidence of nephrotoxicity. Doses of 8 and 10 g/kg produced coagulative necrosis of inner cortical tubules. Cephaloridine (1.1 g/kg) was associated with more severe tubular necrosis than was ceftazidime (10 g/kg), the exerted its toxicity primarily on tubules of the outer cortex. The concurrent administration of furosemide 50 mg/kg potentiated the nephrotoxicity of cephaloridine but not that of ceftazidime. Pre-treatment with probenecid (100 mg/kg) prevented the nephrotoxicity of cephaloridine but not that of ceftazidime.

In male rats, single sc doses of ceftazidime, 4 g/kg or more, produced acute tubular necrosis (inner cortex) and elevations in serum urea nitrogen. This effect was not potentiated by concurrent administration of either gentamicin (35 mg/kg) or furosemide (100 mg/kg), nor was it prevented by pre-treatment with probenecid (100 mg/kg).

In both mice and rats (single dose studies), prolonged observation indicated that the tubular necrosis caused by a single dose of ceftazidime was maximal in severity 48 hours post-dose. Significant improvement was noted after 3 days, with nearly complete regeneration after 7 days.

Male rats given sc injections of ceftazidime 4 g/kg/day for 10 days exhibited increased urinary excretion of enzymes, protein and epithelial cells, which were maximal on day 2 but gradually returned to normal with continued treatment. Ten day treatment with either gentamicin (35 mg/kg/day), amikacin (250 mg/kg/day), or tobramycin (60 mg/kg/day) produced necrosis, mainly of outer cortical tubules. Combination of an aminoglycoside regimen with ceftazidime 4 g/kg/day produced inner cortical tubular necrosis similar to that observed for ceftazidime alone, but with less outer cortical tubular necrosis than that caused by the aminoglycoside alone.

In the rabbit, single ceftazidime doses of 500 mg/kg IM, and 400 or 800 mg/kg SC were not nephrotoxic. Cephaloridine (140 mg/kg IM or 200 mg/kg SC), and cefazolin (800 mg/kg SC) caused marked abnormalities of plasma urea and creatinine, and of tubular ion transport, gluconeogenesis and histology.

### ***Mutagenicity Studies***

Ceftazidime was evaluated *in vivo* and *in vitro* in a series of standard mutagenicity assays.



### ***In Vitro Assays***

The mutagenic potential of ceftazidime was assessed using a modified Ames test, a modified fluctuation test (Harefield) and a yeast gene conversion test (Davis). The results of the Ames plate incorporation assay, in which six concentrations of antibiotic were tested in the presence and absence of microsomes, showed a significantly positive result with *Salmonella typhimurium* strain TA 1537 at 0.9 µg of ceftazidime/plate but this was believed to have occurred by chance. Ceftazidime was negative in the modified Ames test in which it was pre-incubated with liver microsomes.

In the modified pre-incubation fluctuation test, no mutagenic effects were observed at ceftazidime concentrations up to 430 µg/mL.

Similarly, ceftazidime did not induce detectable gene conversion in *Saccharomyces cerevisiae* JD1 cells at concentrations up to 860 µg/mL.

### ***In-vivo Micronucleus Test***

In a micronucleus test, mice received single intraperitoneal injections of 0.56, 1.67 or 5.02 g/kg ceftazidime. No evidence of a clastogenic effect was noted.

### ***In-vivo Cytogenicity Study***

A micronucleus test was used to compare the clastogenic properties of freshly prepared solutions of ceftazidime with samples of ceftazidime stored for up to 24 hours at 25°C. Mice were injected with single IP doses of 1.0 or 2.5 g/kg. Neither fresh nor stored (which contains pyridine as a degradation product of the antibiotic during storage)

solutions of ceftazidime induced a significant increase in detectable chromosomal damage. However, a significant ( $p < 0.05$ ) reduction in the ratio of immature to mature erythrocytes occurred in mice given 2.5 g/kg of ceftazidime (stored solution) 24 hours previously.

### ***Tolerance Studies***

An aqueous solution of ceftazidime 25% w/v was tested for intramuscular irritancy in both adult (1.0 mL) and infant (0.2 mL) rabbits. Lesions consisting of hemorrhage, inflammation, and necrosis were produced which regressed and had almost completely healed by 14 days post-dose.

Intra-arterial injection of 0.5 mL ceftazidime 25% w/v into rabbit ears produced minimal local damage, being morphologically and histologically similar to that caused by intra-arterial injection of 0.5 mL normal saline.

The intracisternal injection of ceftazidime, ampicillin sodium, and gentamicin sulphate caused convulsions of dose-related severity in male rabbits. The minimum dosage levels at which convulsions were observed were 5 mg/kg for ceftazidime, 12 mg/kg for ampicillin sodium, 4.5 mg/kg for gentamicin sulphate.

### ***Immunological Studies***

Ceftazidime (25 mg/kg, IM) was administered as an aqueous solution to 10 rabbits (5M, 5F) once weekly for 6 weeks. Sera taken 7 days after the last dose were negative for ceftazidime antibody by both enzyme-linked immunosorbent assay (ELISA) and passive cutaneous anaphylaxis (PCA) test.

Four doses of ceftazidime (25 mg/kg) in an aqueous emulsion with Freund's adjuvant were given to 6 rabbits (3M, 3F) as a single inoculation followed by boosters on days 21, 56 and 95. Sera taken on day 102 were all negative for drug antibody by ELISA, but 1 out of 6 (female subject) was positive by PCA test. This antibody was skin fixing and heat labile.

Antisera prepared with cephaloridine, cephalexin, cephalothin, and cefotaxime showed cross-reactivity with a ceftazidime-cytochrome C antiserum, but did not cross-react with an antiserum prepared with a ceftazidime-human gamma globulin conjugate.

The effects of storage of ceftazidime (either as a dry powder at 37°C for 4 months or as a 25% w/v solution at 25°C for 72 hours) on its immunogenicity and elicitoricity (i.e. the ability to produce anaphylaxis in an immunized subject) were studied in rabbits and guinea pigs. Immunogenicity was unaffected but elicitoricity was found to increase with storage.

Solutions of ceftazidime and its degradation products, formed on both wet and dry storage as above, did not cause the release of allergic mediators from human peripheral blood basophils or from fragments of human lung parenchyma *in vitro*.

## **Reproduction and Teratology Studies**

### Teratology

#### Mouse

Four groups of pregnant female mice were administered sc injections of either saline (28 mice) or ceftazidime (1.5 g/kg/day - 21 mice, 3.25 g/kg/day - 20 mice, 6.5 g/kg/day - 29 mice) from day 6 to day 15 of pregnancy inclusive (period of organogenesis). Eight mice from the control group and eight from the group given 6.5 g/kg/day were allowed to

give birth and rear their young to weaning. The remaining animals were sacrificed on day 18 of pregnancy and an examination made of their uterine contents.

The following external or soft tissue defects were found, each occurring in a single fetus only: control - left testis absent (1 mouse); 1.5 g/kg - small depression in palate (2 mice), right testis not found (1 mouse); cleft palate (1 mouse); 3.25 g/kg - small swelling at base of tail (1 mouse); 6.5 g/kg - enlarged space in thoracic cavity (1 mouse); cleft palate (1 mouse), enlarged thin walled bladder (1 mouse), small depression in palate (1 mouse).

The high incidence of skeletal variants seen in all groups (control: 39.51%, 1.5 g/kg: 53.98%, 3.25 g/kg: 50.70%, and 6.5 g/kg: 63.55%) was due to the large number of fetuses with supernumerary ribs. The incidence of rib variants was significantly higher ( $p < 0.05$ ) in the high-dose group (6.5 g/kg) than in the control group.

The overall incidence of skeletal abnormalities was 15% (controls), 20% (3.25 g/kg ceftazidime) and 24% (6.5 g/kg ceftazidime). These consisted mainly of obliquely fused sternbrae. In the group treated with the high dose (6.5 g/kg), one fetus had extra ribs on cervical vertebrae 6 and 7 and one fetus had a bifid hyoid bone.

The number of live pups/litter born to mice treated with the high-dose (6.5 g/kg) was significantly ( $p < 0.05$ ) lower (10) when compared to controls (13). Similarly the litter weights for the treated group were consistently and significantly ( $p < 0.05$ ) lower than those in the control group throughout lactation.

### Rabbit

Female Dutch rabbits were given intramuscular injections of 0 (18 rabbits), 25 mg/kg (27), 50 mg/kg (18), 100 mg/kg (18) or 200 mg/kg (9) ceftazidime daily from day 6 to day

18 of pregnancy (organogenesis). On day 29, the rabbits were sacrificed and the uterine contents examined.

Twenty-nine rabbits dosed with ceftazidime were either found dead (18) or had to be destroyed (11) due to ill health (diarrhea and emaciation) or because they had aborted their fetuses. One rabbit in the control group was found dead on day 10 of pregnancy. The incidence of death was not dose-related (highest incidence occurred in the group given 25 mg/kg/day).

A decrease in body weight was noted during the first week of dosing and continued for the duration of the study in those rabbits receiving doses greater than 25 mg/kg of ceftazidime per day.

Results of the examination of the uterine contents are presented in the Table 18.

**TABLE 18**

Observation	MEAN RESULTS				
	Control	25 mg/kg ceftazidime	50 mg/kg ceftazidime	100 mg/kg ceftazidime	200 mg/kg ceftazidime
Implantations	7	6	6	6	6
Resorptions	1	1	2	2	4
Live Fetuses	6	5	4	4	3
Live Litter Weight (g)	191	153	136	141	138
Within Litter Mean Live Fetus Weight (g)	31.4	30.2	28.6	26.9	24.5
Within Litter Mean Placenta Weight (g)	3.93	4.56	3.56	3.87	2.91

Two dead fetuses were reported - one in the control group (flexed forepaws) and one in 25 mg/kg/day group. Three fetuses (25 mg/kg group) from a litter of 5 had one or more of the following gross external abnormalities: anencephaly, gastroschisis, 1st and 3rd toes absent from both forepaws, 4th toe on right hind paw absent, tail twisted,

craniorachischisis, lower jaw absent, eyes open, fore and hind limb buds present, tail and anogenital papilla present, thoracic and abdominal organs exposed.

### ***Peri- and Postnatal Study***

Groups of 20 female AHA rats (approximately 10 wk of age, 200 g body weight) received a daily SC injection of either 0, 0.1, 0.5 or 2.5 g/kg ceftazidime. Animals were dosed from day 17 of pregnancy to the day of parturition and subsequently on days 1-21 inclusive postpartum.

No significant adverse reactions were seen during pregnancy with the exception of the high dose (2.5 g/kg) group which produced large quantities of soft wet feces. During the second and third week of the lactation period the dams treated with ceftazidime gained weight more rapidly than the control group and this effect was dose-related.

At termination (day 21 postpartum), pups in the high dose group (2.5 g/kg) had gained significantly ( $p < 0.05$ ) less weight (47.95 g) than controls (52.23 g). This was observed throughout lactation. Two of the dams in the high dose (2.5 g/kg) group were killed due to the death of their litters. Both animals had gastrointestinal disorders due to heavy growth of Gram-positive *Streptococcus*. One dam in the 0.5 g/kg group was killed due to ill health (diarrhea due to bacterial typhlitis).

### ***Fertility and Reproduction***

Groups of 20 male and 40 female mice received SC injections of either saline or 1.5, 3.25 or 6.5 g/kg of ceftazidime daily throughout gametogenesis and mating and in the case of females through pregnancy. Males were treated for 60 days prior to mating and females for 14 days. One half of the pregnant mice were sacrificed on day 18 of pregnancy while the remainder were allowed to litter and rear their young for 21 days.

Two pups from each litter were retained to study any effects on fertility of the F<sub>1</sub> generation.

Treatment with ceftazidime had no adverse effect on the fertility of either male or female mice.

A high incidence of skeletal variants seen in all of the groups (control: 45.48%, 1.5 g/kg/day: 55.04%, 3.25 g/kg/day: 64.40%, 6.5 g/kg/day: 73.97%) was due to the large number of fetuses with supernumerary ribs.

The incidence of bone variants was significantly higher ( $p < 0.05$ ) in the high-dose group (6.5 g/kg/day) as compared to the controls. Throughout lactation, the mean pup weights (F<sub>1</sub> generation) for the mid (3.25 g/kg/day) and high (6.5 g/kg/day) dose groups were lower than the corresponding control values but the differences did not achieve statistical significance.

There were no significant differences in pregnancy rates for any of the F<sub>1</sub> generation groups.

The mean pup weights (F<sub>2</sub> generation) during lactation in the high-dose group were consistently less than those of controls but the differences were not statistically significant and this was attributed to the lighter weights of the dams.

## References or Selected Bibliography

1. Ackerman BH, Ross J, Tofte W, Rotschafer JC. Effect of decreased renal function on the pharmacokinetics of ceftazidime. *Antimicrobial Agents and Chemotherapy* 1984; 25(6):785-786.
2. Acred P, Ryan DM, Sowa MA, Watts CM. The in vitro antibacterial activity of ceftazidime (GR 20263) - a comparison with other new beta-lactam antibiotics and gentamicin. *Journal of Antimicrobial Chemotherapy* 1981; 8(Suppl B):247-255.
3. Adam D, Reichart B, Williams KJ. Penetration of ceftazidime into human tissue in patients undergoing cardiac surgery. *Journal of Antimicrobial Chemotherapy* 1983; 12(Suppl.A):269-273.
4. Assael BM, Boccazzi A, Caccamo ML, Giunta A, Marini A, Padoan R et al. Clinical Pharmacology of Ceftazidime in Paediatrics. *J Antimicrob Chemother* 1983; 12(SUPPL A):341-346.
5. Bauernfeind A. An evaluation of the activity of cephalosporins against *Pseudomonas aeruginosa*. *Journal of Antimicrobial Chemotherapy* 1981; 8(Suppl.B):111-117.
6. Bint AJ, Yeoman P, Kilburn P, Anderson R, Stansfield E. The in vitro activity of ceftazidime compared with that of other cephalosporins. *Journal of Antimicrobial Chemotherapy* 1981; 8(Suppl.B):47-51.
7. Brumfitt W, Hamilton-Miller JMT. The susceptibility of nosocomial pathogens to ceftazidime. *Journal of Antimicrobial Chemotherapy* 1981; 8(Suppl B):15-21.
8. Chow AW, Bartlett KH. Comparative in vitro activity of ceftazidime (GR 20263) and other beta-lactamase stable cephalosporins against anaerobic bacteria. *Journal of Antimicrobial Chemotherapy* 1981; 8(Suppl. B):91-95.
9. Chow AW, Bartlett KH. Comparative in vitro activity of ceftazidime (GR 20263) and other beta-lactamase stable cephalosporins against *Pseudomonas*. *Journal of Antimicrobial Chemotherapy* 1981; 8(Suppl B):345-348.
10. Clumeck N, Gordts B, Dab I, Jaspar N, Van Laethem Y, Butzler JP. Ceftazidime as a single agent in the treatment of severe *Pseudomonas aeruginosa* infections. *Journal of Antimicrobial Chemotherapy* 1983; 12(Suppl A):207-211.
11. Clumeck N, Van Laethem Y, Gordts B, Jaspar N, Butzler JP. Use of ceftazidime in the therapy of serious infections, including those due to multiresistant organisms. *Antimicrobial Agents and Chemotherapy* 1983; 24(2):176-180.



12. Cox CE. A comparison of ceftazidime and tobramycin in the treatment of complicated urinary tract infections. *Journal of Antimicrobial Chemotherapy* 1983; 12(Suppl.A):47-52.
13. Daschner FD, Petersen EE, Just HM, Hillemanns HG. Penetration of Ceftazidime Into Serum, Myometrium, Endometrium, Salpinges and Subcutaneous Tissue. *J Antimicrob Chemother* 1983; 12(SUPPL. A):247-249.
14. Davies BI, Maesen FPV, van Noord JA. Treatment of chronic and recurrent respiratory infections with intramuscular ceftazidime. *Journal of Antimicrobial Chemotherapy* 1983; 12(Suppl.A):1-8.
15. Dutoy JP, Wauters G. The treatment of bone infection with ceftazidime. *Journal of Antimicrobial Chemotherapy* 1983; 12(Suppl.A):229-233.
16. Gooch WM, Swenson E. Use of ceftazidime in the management of bacterial cellulitis in infants and children. *Current Therapeutic Research* 1985; 37(1):3-8.
17. Harding SM, Harper PB. The pharmacokinetic behaviour of ceftazidime in man and the relationship between serum levels and the in vitro susceptibility of clinical isolates. *Infection* 1983; 11(Suppl.1):S49-S53.
18. Harper PB. The in vitro properties of ceftazidime. *Journal of Antimicrobial Chemotherapy* 1981; 8(Suppl.B):5-13.
19. Hayes MV, Orr DC. Mode of action of ceftazidime: affinity for the penicillin-binding proteins of *Escherichia coli* K12, *Pseudomonas aeruginosa* and *Staphylococcus aureus*. *Journal of Antimicrobial Chemotherapy* 1983; 12: 119-126.:119-126.
20. Heintz, 2009. Antimicrobial dosing concepts and recommendations for critically ill adult patients receiving continuous renal replacement therapy or intermittent hemodialysis. *Pharmacotherapy*, 2009; 29 (5):562-577
21. Jones RN, Barry AL, Thornsberry C, Gerlach EH, Fuchs PC, Gavan TL et al. Ceftazidime, a *Pseudomonas* active cephalosporin; in vitro antimicrobial activity evaluation including recommendations for disc diffusion susceptibility tests. *Journal of Antimicrobial Chemotherapy* 1981; 8(Suppl.B):187-211.
22. Kemmerich B, Warns H, Lode H, Borner K, Koeppe P, Knothe H. Multiple-dose pharmacokinetics of ceftazidime and its influence on fecal flora. *Antimicrobial Agents and Chemotherapy* 1983; 24(3):333-338.
23. Leeder JS, Spino M, Isles AF, Tesoro AM, Gold R, MacLeod SM. Ceftazidime disposition in acute and stable cystic fibrosis. *Clinical Pharmacology and Therapeutics* 1984; 36(3):355-362.

24. Low DC, Bissenden JG, Wise R. Ceftazidime in neonatal infections. *Archives of Disease in Childhood* 1985; 60: 360-364.:360-364.
25. Lundbergh P, Jarstrand C, Morfeldt-Manson L, Weiland O. Ceftazidime in septicaemia. *Journal of Antimicrobial Chemotherapy* 1983; 12(Suppl.A):199-205.
26. Mandell LA, Nicolle LE, Ronald AR, Duperval R, Robson HG, Feld R et al. A multicentre prospective randomized trial comparing ceftazidime with cefazolin/tobramycin in the treatment of hospitalized patients with non-pneumococcal pneumonia. *Journal of Antimicrobial Chemotherapy* 1983; 12(Suppl.A):9-20.
27. Matzke G et al. Determinants of ceftazidime clearance by continuous venovenous hemofiltration and continuous venovenous hemodialysis. *Antimicrob Agents and Chemo*, 2000; 44:1639-1644
28. Modai J, Vittecoq D, Decazes JM, Wolff M, Meulemans A. Penetration of ceftazidime into cerebrospinal fluid of patients with bacterial meningitis. *Antimicrobial Agents and Chemotherapy* 1983; 24(1):126-128.
29. Mulhall A, de Louvois J. The pharmacokinetics and safety of ceftazidime in the neonate. *Journal of Antimicrobial Chemotherapy* 1985; 15(1):97-103.
30. Naber KG, Kees F, Grobecker H. Ceftazidime: pharmacokinetics in young volunteers versus elderly patients and therapeutic efficacy with complicated urinary tract infections. *Journal of Antimicrobial Chemotherapy* 1983; 12(Suppl.A):41-45.
31. Neu HC, Labthavikul P. Antibacterial activity and beta lactamase stability of ceftazidime, an aminothiazolyl cephalosporin potentially active against *Pseudomonas aeruginosa*. *Antimicrobial Agents and Chemotherapy* 1982; 21(1):11-18.
32. Norrby SR, Burman LA, Linderholm H, Trollfors B. Ceftazidime: pharmacokinetics in patients and effects on the renal function. *Journal of Antimicrobial Chemotherapy* 1982; 10(3):199-206.
33. Piot P, Van Dyck E, Colaert J. In vitro activity of ceftazidime (GR 20263) and other beta lactam antibiotics against *Haemophilus influenzae*. *Infection* 1983; 11(Suppl.1):S32-S34.
34. Pollock I, Mulhall A, de Louvois J. Ceftazidime in the treatment of neonatal infection. *Journal of Hospital Infection* 1985; 6:158-165.

35. Reed MD, O'Brien CA, Aronoff SC, Klinger JD, Blumer JL. Ceftazidime as initial therapy for suspected bacterial infections in hospitalized pediatric patients. *Antimicrobial Agents and Chemotherapy* 1984; 26(3):318-321.
36. Richards DM, Brogden RN. Ceftazidime. A review of its antibacterial activity, pharmacokinetic properties and therapeutic use. *Drugs* 1985; 29:105-161.
37. Shah PM. Bactericidal activity of ceftazidime against *Pseudomonas aeruginosa* under conditions simulating serum pharmacokinetics. *Journal of Antimicrobial Chemotherapy* 1981; 8(Suppl.B):135-140.
38. Simpson IN, Plested SJ, Harper PB. Investigation of the beta-lactamase stability of ceftazidime and eight other new cephalosporin antibiotics. *Journal of Antimicrobial Chemotherapy* 1982; 9: 357-368:357-368.
39. Sommers DK, Walters L, Van Wyk M, Harding SM, Paton AM, Ayrton J. Pharmacokinetics of ceftazidime in male and female volunteers. *Antimicrobial Agents and Chemotherapy* 1983; 23(6):892-896.
40. Thabaut A, Durosoir JL, Saliou P. Comparison of the in vitro activities of ceftazidime and new cephalosporins against 107 strains of *Pseudomonas aeruginosa* and 249 strains of cefazolin-resistant Enterobacteriaceae. *Journal of Antimicrobial Chemotherapy* 1981; 8(Suppl.B):123-125.
41. Tourkantonis A, Nicolaidis P. Pharmacokinetics of ceftazidime in patients undergoing peritoneal dialysis. *Journal of Antimicrobial Chemotherapy* 1983; 12(Suppl.A):263-267.
42. Turner A, Pedler SJ, Carswell F, Spencer GR, Speller DCE. Serum and sputum concentrations of ceftazidime in patients with cystic fibrosis. *Journal of Antimicrobial Chemotherapy* 1984; 14(5):521-527.
43. Walstad RA, Thurmann-Nielson E, Dale LG, Brunn JN. Blister and lymphatic fluid studies of ceftazidime, a new cephalosporin with antipseudomonal activity. *British Journal of Clinical Pharmacology* 1982; 14(4):626P.
44. Walstad RA, Hellum KB, Blika S, Dale LG, Fredriksen T, Myhre KI et al. Pharmacokinetics and tissue penetration of ceftazidime: studies on lymph, aqueous humour, skin blister, cerebrospinal and pleural fluid. *Journal of Antimicrobial Chemotherapy* 1983; 12(Suppl.A):275-282.
45. Welage LS, Schultz RW, Schentag JJ. Pharmacokinetics of ceftazidime in patients with renal insufficiency. *Antimicrobial Agents and Chemotherapy* 1984; 25(2):201-204.

46. Wittman DH, Schassan HH. Penetration of eight beta-lactam antibiotics into the peritoneal fluid. Archives of Surgery 1983; 118:205-213.
47. Wittmann DH, Schassan HH, Kohler F, Seibert W. Pharmacokinetic studies of ceftazidime in serum, bone, bile, tissue fluid and peritoneal fluid. Journal of Antimicrobial Chemotherapy 1981; 8(Suppl.B):293-297.
48. Clinical and Laboratory Standards Institute (CLSI). Performance Standards for Antimicrobial Susceptibility Testing; Twenty-fourth Informational Supplement. 2014. CLSI document M100-S24, Clinical and Laboratory Standards Institute, 950 West Valley Road, Suite 2500, Wayne, Pennsylvania 19087, USA